

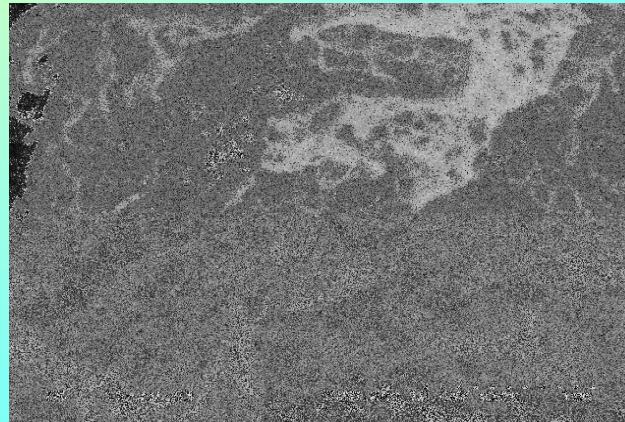
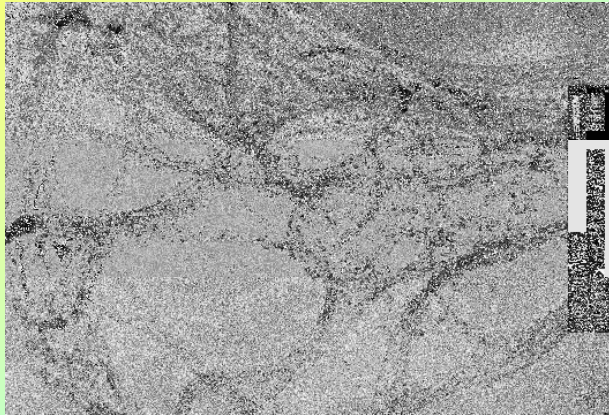


**Valerian E. Kagan**

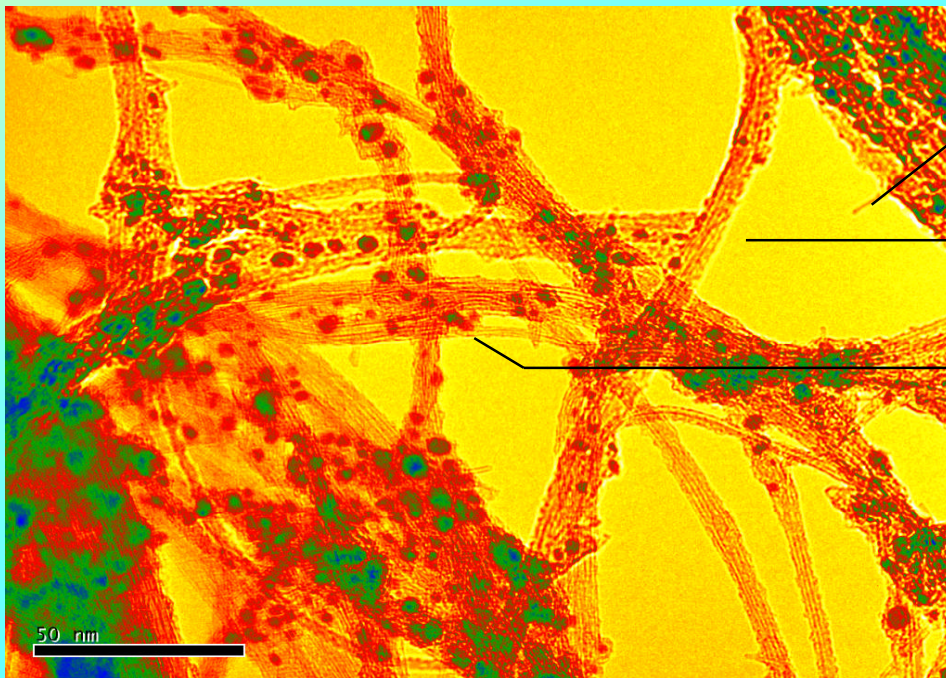
**Macrophage Response to Single Walled Carbon Nanotubes:  
Oxidative Stress and Inflammatory Consequences.**

**Center For Free Radical and Antioxidant Health, Department of Environmental and  
Occupational Health, University of Pittsburgh,**

# Single Walled Carbon Nanotubes



**'Tangles' of nanotubes and nanoropes**



**Nanotubes**

**Nanoropes**

**Catalyst particles**

**Raw single walled carbon nanotube material.**

**Handling nanotube material**



**Raw SWCNT material**

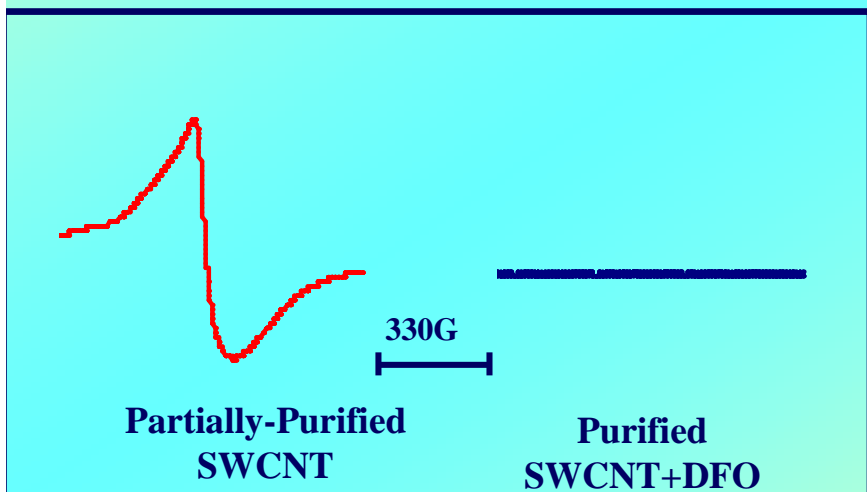
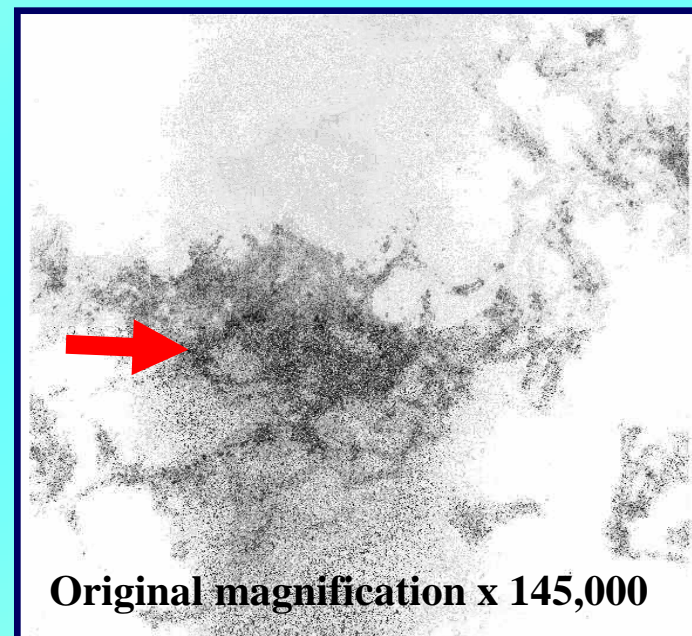
*Courtesy of Andrew Maynard*



## Metal Components of SWCNT

Component	( $\mu\text{g}/\text{gram}$ )	Component	( $\mu\text{g}/\text{gram}$ )
Aluminum	233	Molybdenum	1070
Calcium	164	Sodium	8750
Cadmium	23.4	Nickel	8,750
Chromium	13.1	Palladium	28
Copper	2,530	Selenium	<2.001
<b>Iron</b>	<b>239,000</b>	Titanium	6.92
		Zinc	85.9

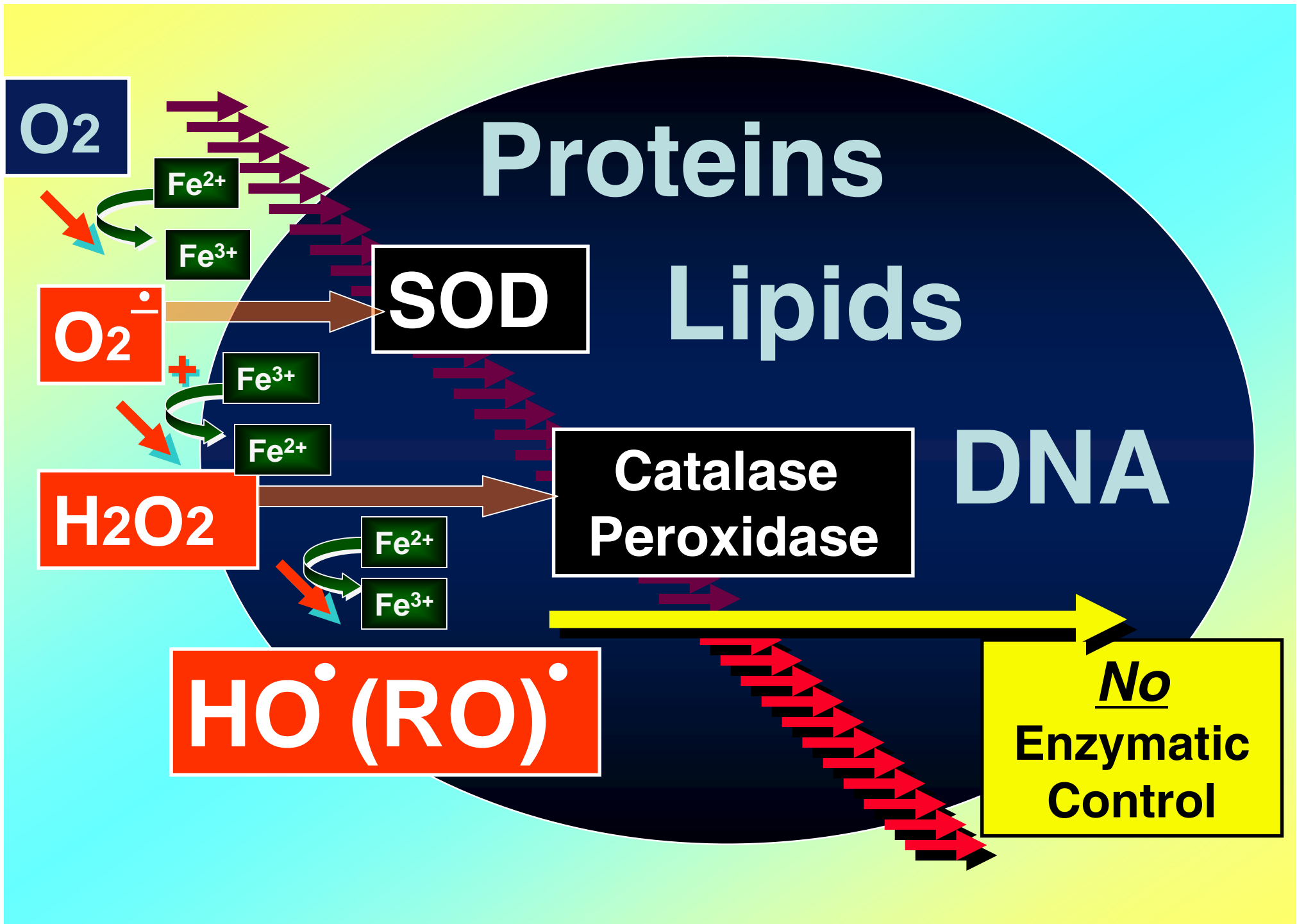
## Transmission Electron Microscopy of SWCNT



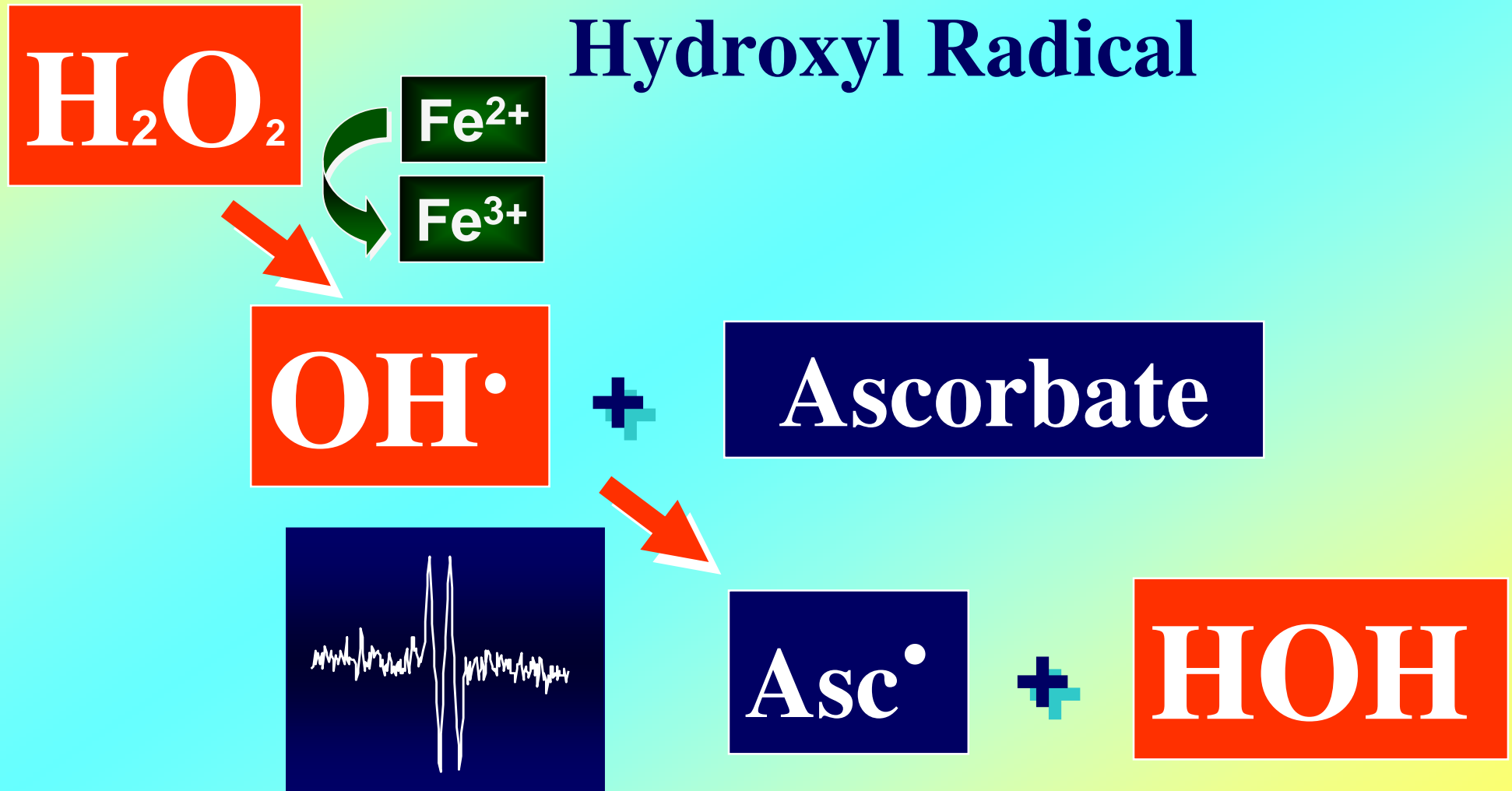
**EPR spectra of partially-purified SWCNT manufactured by high-pressure CO conversion (HiPco™) technology as compared to purified SWCNT additionally treated with an iron chelator, deferoxamine (DFO).**

Note that partially-purified SWCNT displayed a broad signal with g value 2.0 and half-width of 640G, the signal was not detectable in purified DFO-treated SWCNT.

**Raw Samples of  
Carbon  
Nanotubes  
Contain  
Redox-Active  
Iron**



# Iron-Catalyzed Decomposition of $\text{H}_2\text{O}_2$ Forms A Potent Oxidant – Hydroxyl Radical



# EPR spectra of ascorbate radicals generated by partially purified SWCNT.

Ascorbate



Ascorbate+partially purified  
SWCNT,  
5 min



Ascorbate+partially  
purified SWCNT+DFO



Ascorbate+partially  
purified SWCNT +  
DFO, 5 min

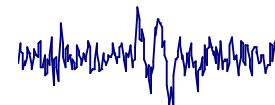


20 G

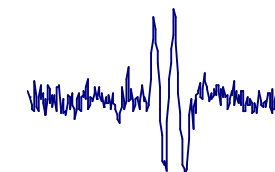
## Model system

**Conditions:** Ascorbate (10 mM) in PBS (pH 7.4); partially purified SWCNT (0.12 mg/ml, 2.5 wt% of iron); desferioxamine, DFO (0.2 mM);

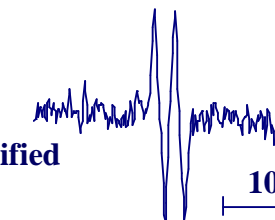
Ascorbate



RAW264.7+  
Ascorbate



RAW264.7 +  
Ascorbate+partially purified  
SWCNT, 5 min



10 G

## RAW264.7 macrophages

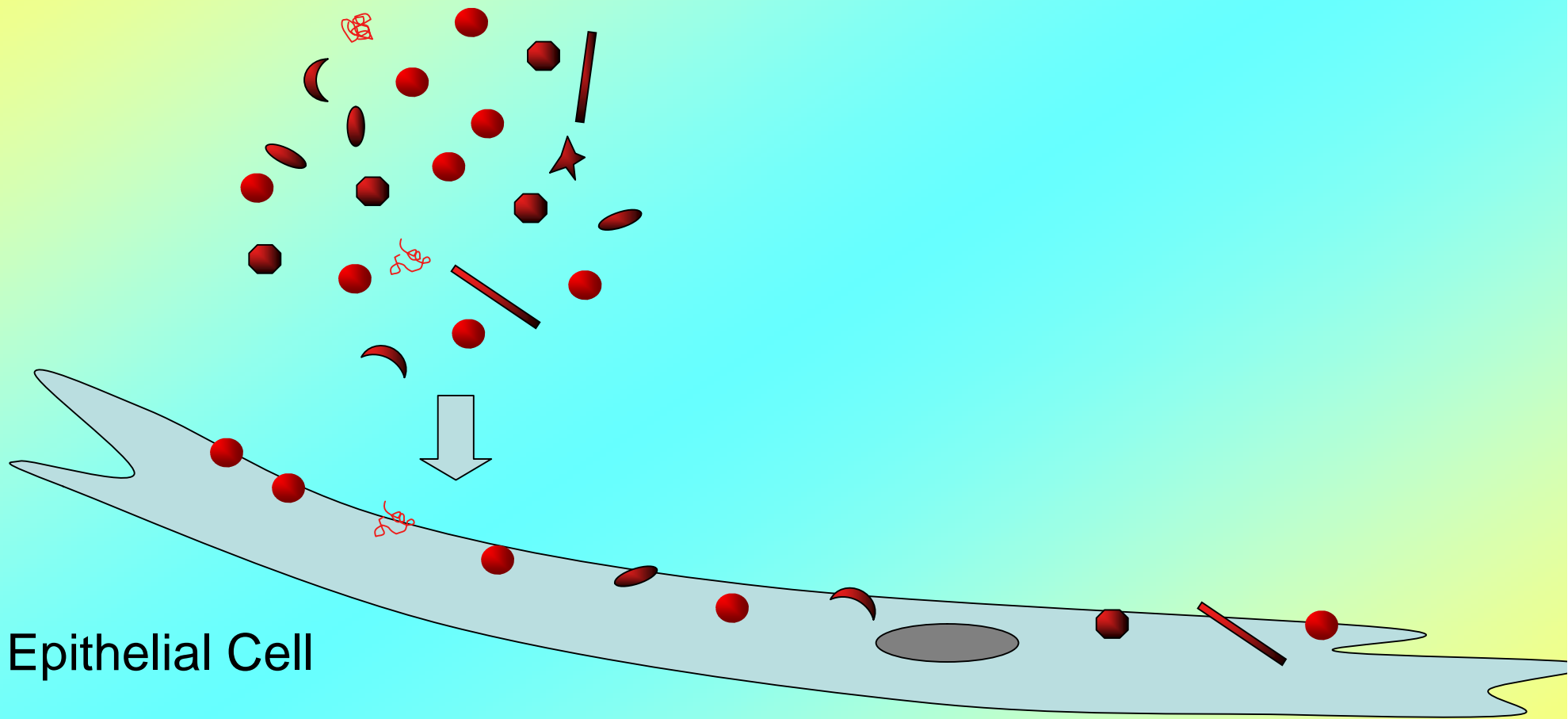
**Conditions:** Zymosan (2.5 mg/ml)-stimulated RAW264.7 macrophages (20x10<sup>6</sup> cells/ml); partially purified SWCNT (2.5 wt% of iron, 0.12 mg/ml)

EPR conditions: microwave power, 20 mW; modulation amplitude, 1.0 G; time constant, 1.3 sec; conversion time, 0.6 sec.

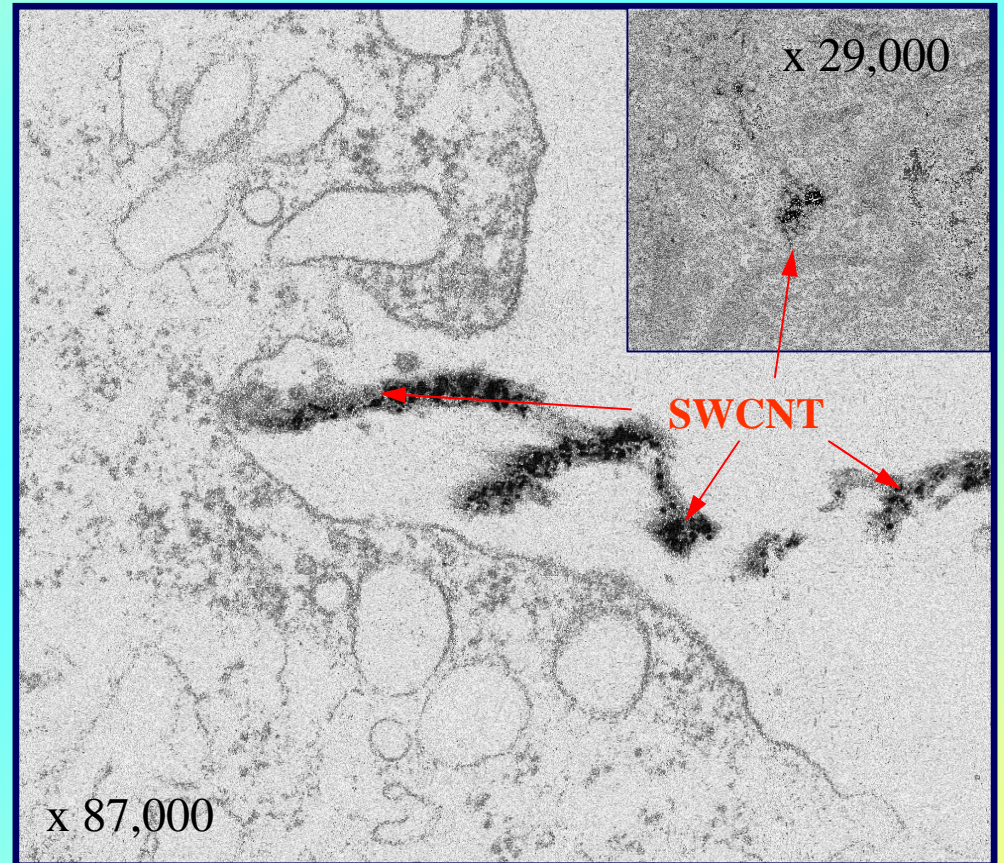
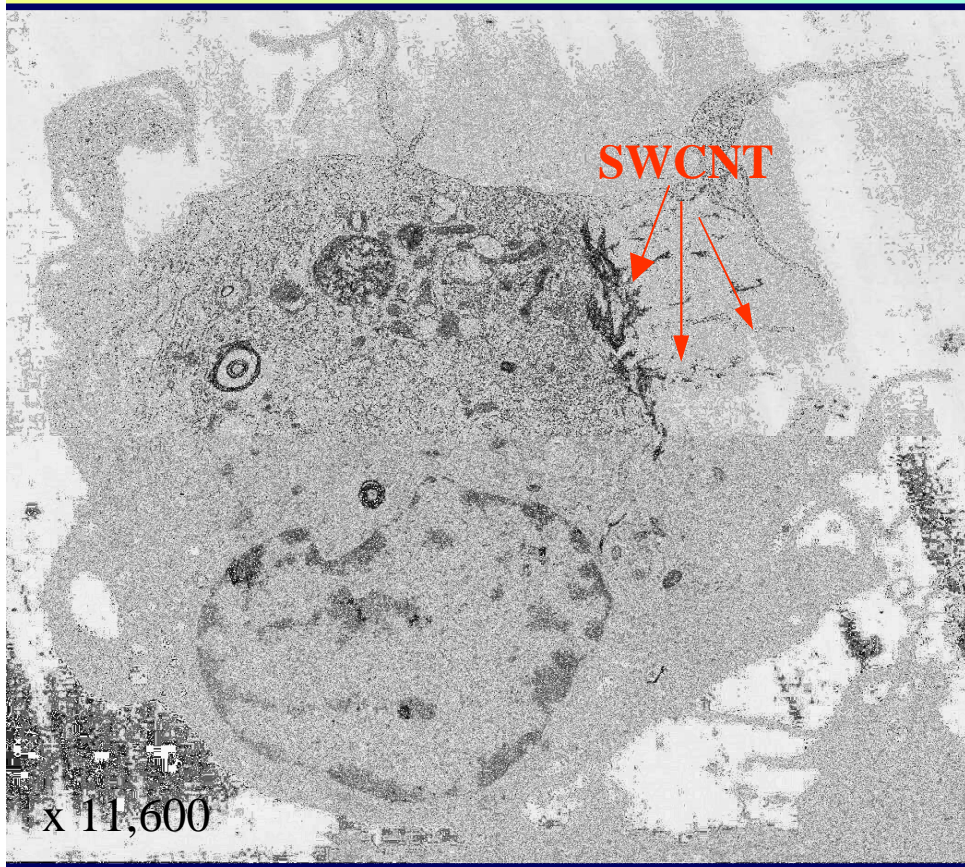
**Carbon  
Nanotubes  
Directly  
Damage  
Broncho-Epithelial  
Cells**



# Particles, Nanotubes



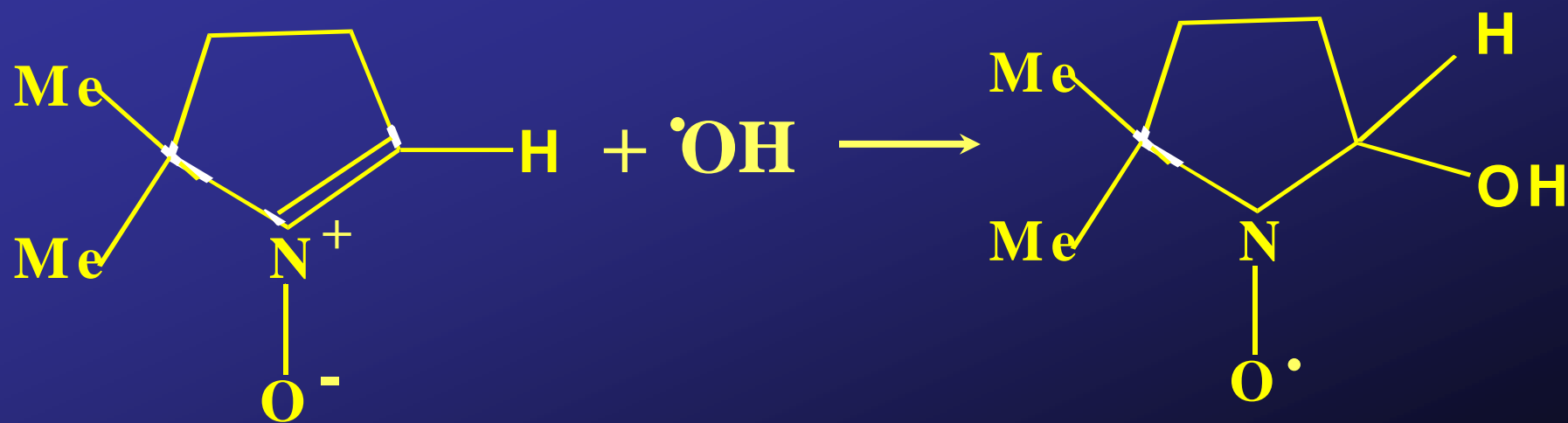
# Engulfment of SWCNT by BEAS-2B Cells



## Fenton Reaction



## DMPO Adduct Formation



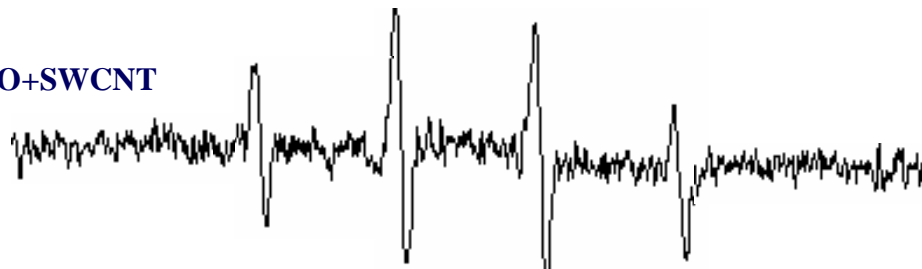
5,5-Dimethyl-1-Pyrroline-N-Oxide  
(DMPO)



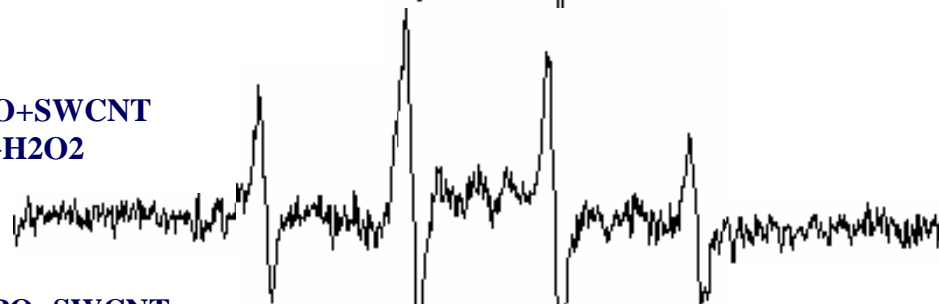
**DMPO**



**DMPO+SWCNT**



**DMPO+SWCNT  
+H<sub>2</sub>O<sub>2</sub>**



**DMPO+SWCNT+  
H<sub>2</sub>O<sub>2</sub>+Catalase**



**DMPO+SWCNT  
+DTPA**



**20 Gauss**

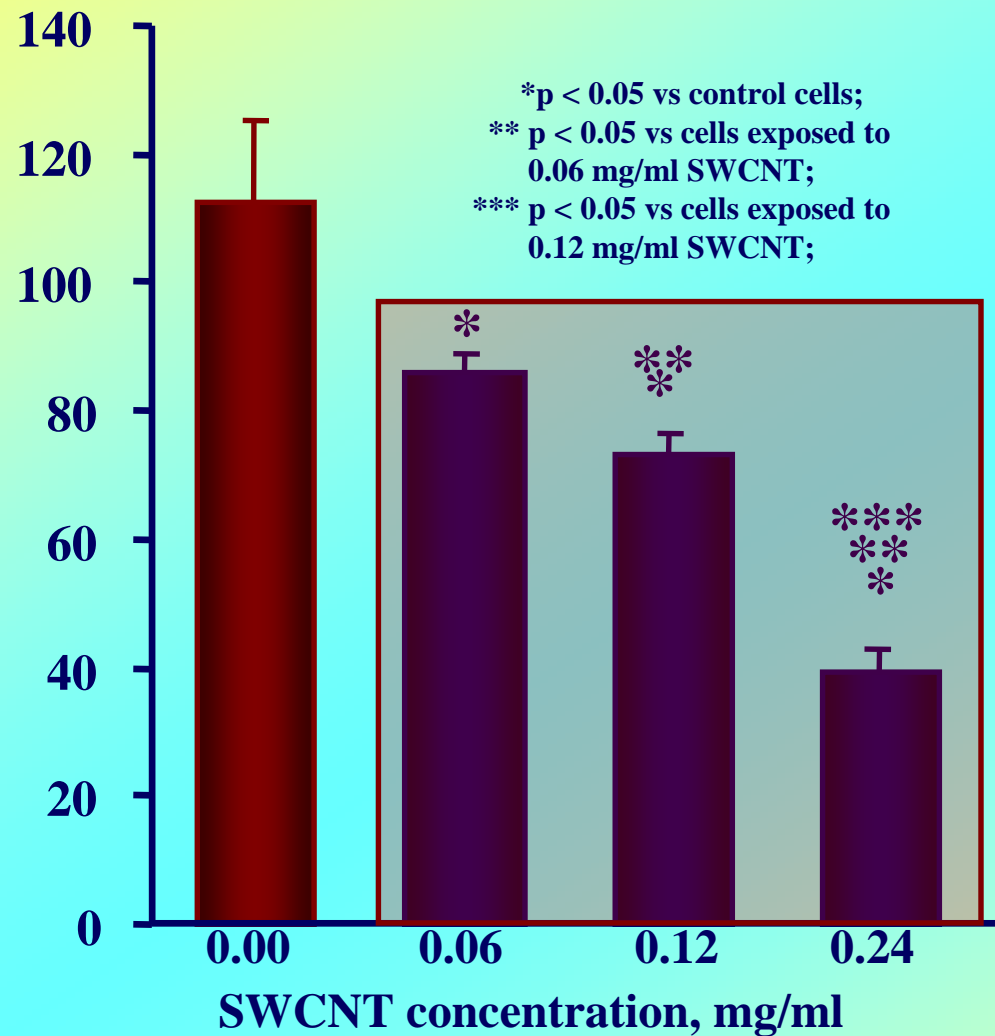
## **ESR spectra of DMPO adducts of free radicals formed by partially purified SWCNT in the presence of BEAS-2B cells.**

**Conditions:** BEAS-2B (2x10<sup>5</sup> cells/ml) in PBS (pH 7.4); 100 mM DMPO; partially purified SWCNT (2.5 wt% of iron, 0.12 mg/ml) H<sub>2</sub>O<sub>2</sub> (1 mM); catalase (20 U/ml); DTPA (0.2 mM).

**ESR conditions:** microwave power, 20 mW; modulation amplitude, 1.0 G; time constant, 1.3 sec; conversion time, 0.6 sec.

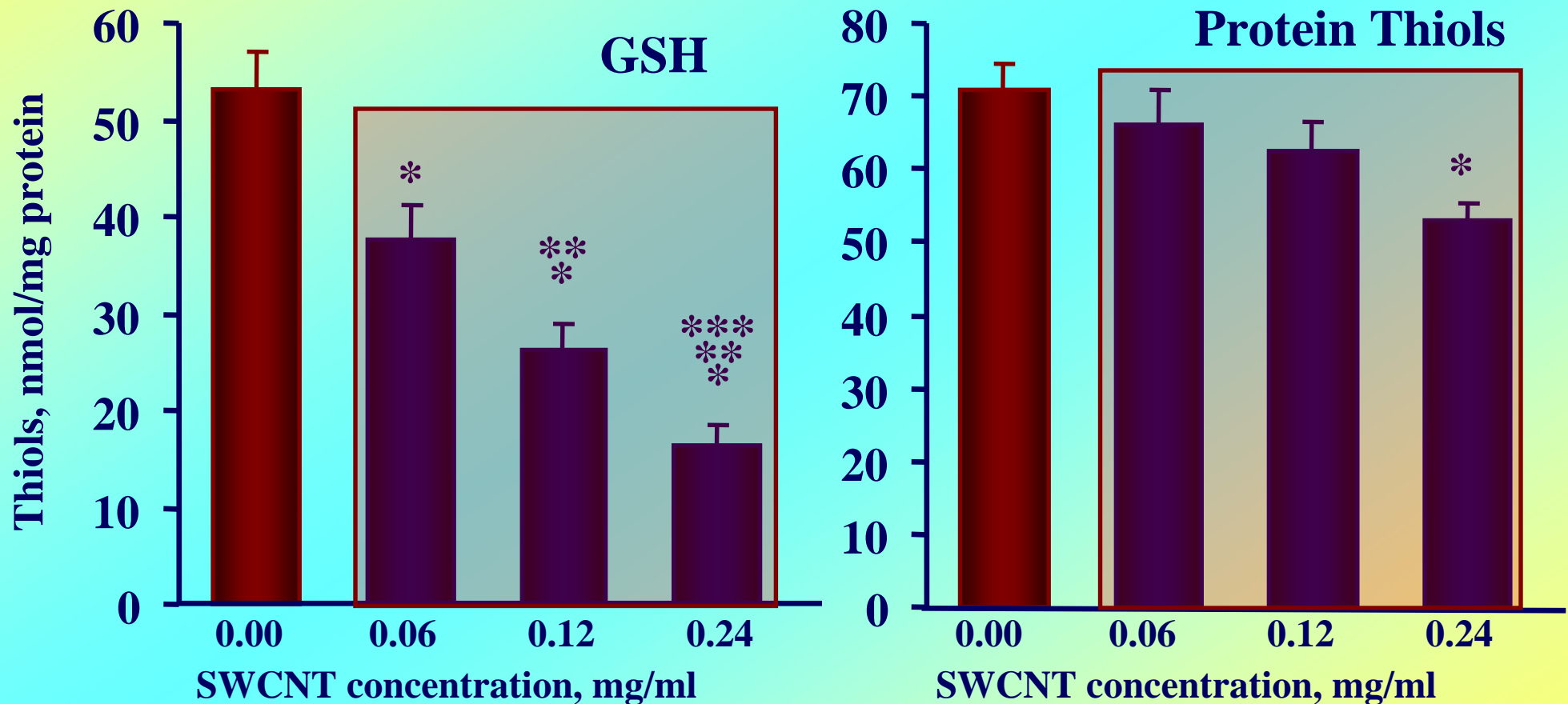


Peroxyl radicals scavenged by  
cells homogenates, nmol/mg protein



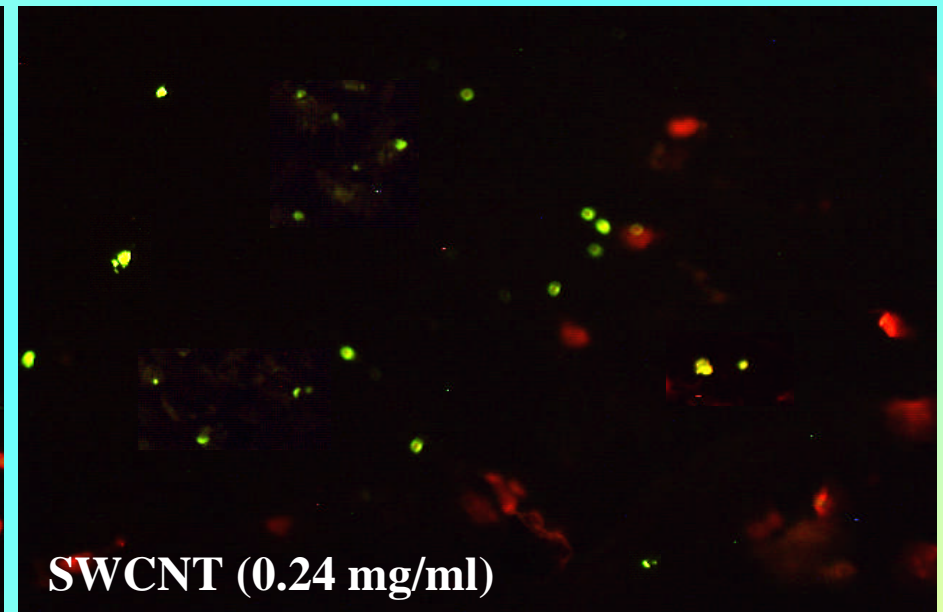
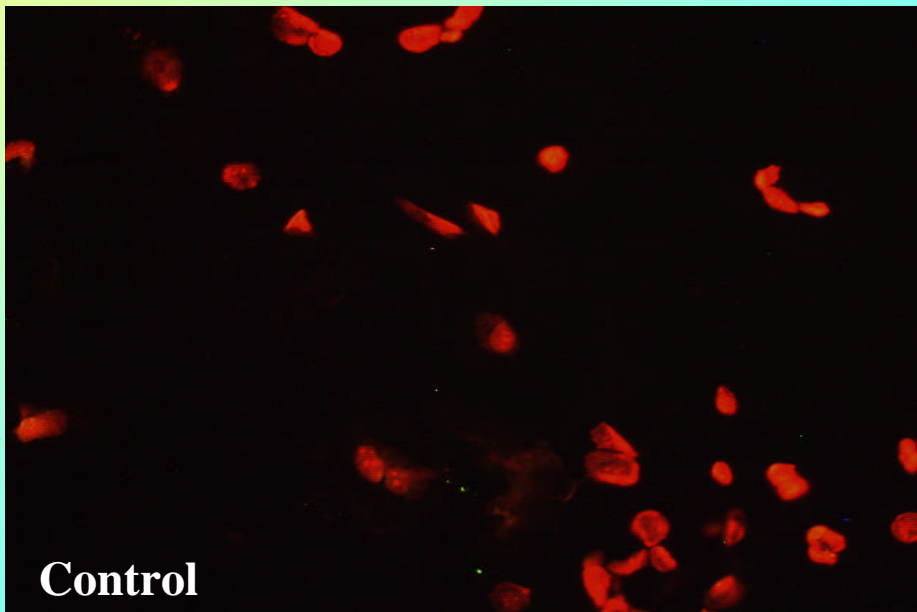
**Total  
Antioxidant  
Reserve in  
BEAS-2B Cells  
Following  
Exposure to  
SWCNT**

# GSH and Protein Thiols in BEAS-2B Cells Following Exposure to SWCNT

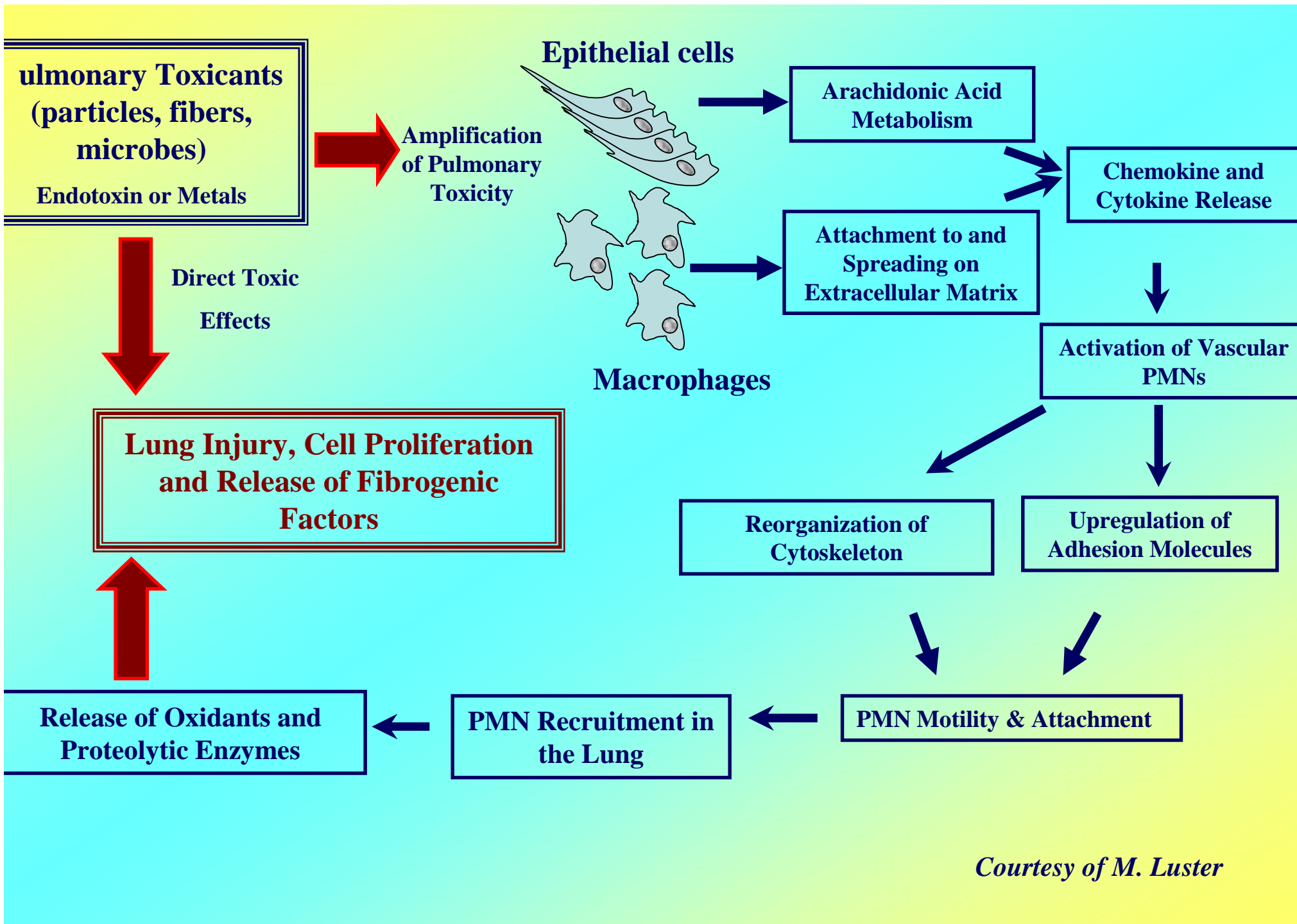


\*p < 0.05 vs control cells; \*\* p < 0.05 vs cells exposed to 0.06 mg/ml SWCNT; \*\*\* p < 0.05 vs cells exposed to 0.12 mg/ml SWCNT;

# Exposure to SWCNT Induced Apoptotic Cell Death



BEAS-2B cells were stained with TUNEL reagent. Apoptotic cells exhibited yellow-green fluorescence, while normal cells counter-stained with propidium iodide fluoresced red. Original magnification x 40.







**Day 0**



**Aspiration with:**  
CNT (10, 20, 40  $\mu\text{g}/\text{mouse}$ )  
UFCB (40  $\mu\text{g}/\text{mouse}$ )  
Silica (40  $\mu\text{g}/\text{mouse}$  & 2.5 mg/mouse)

**Days post Exposure**

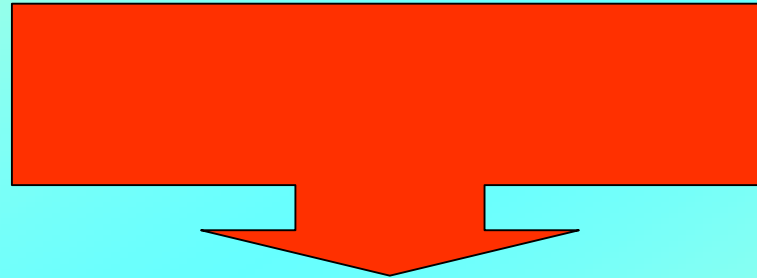
1

3

7

28

60

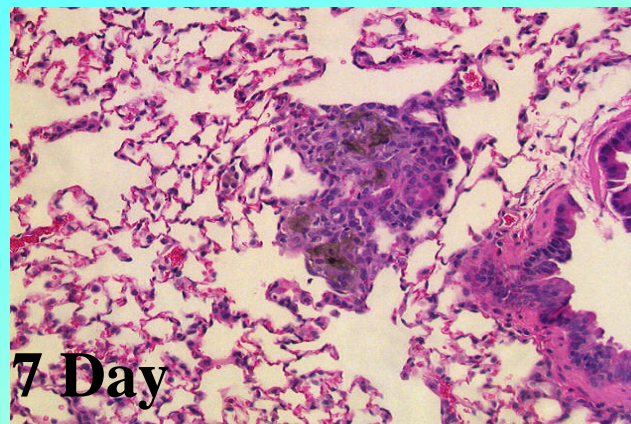
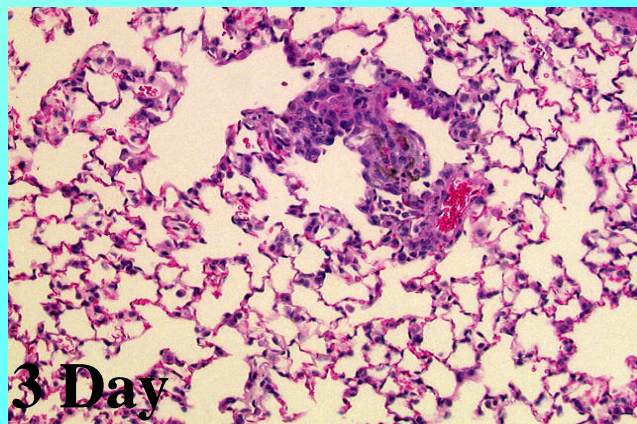
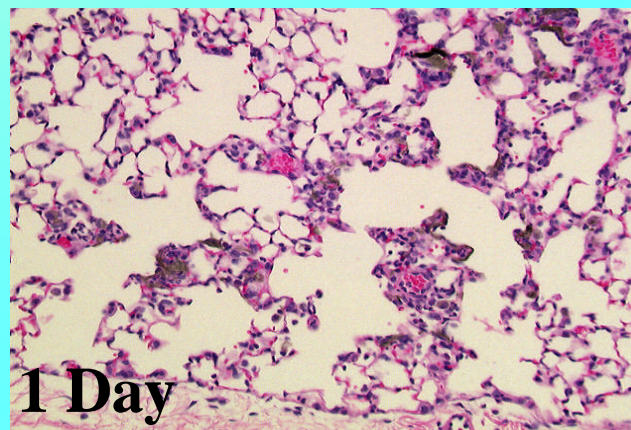
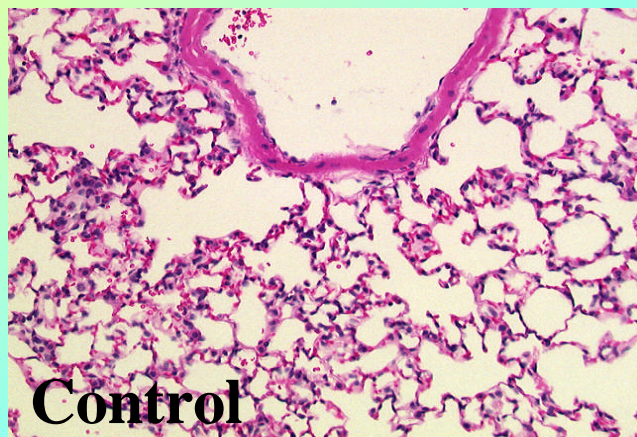


- **Histopathology**
- **Pulmonary injury**
- **Lung functions**
- **Collagen morphometry**
- **Oxidative stress**
- **Cytokines**

**Single Walled Carbon Nanotube Toxicity - purified SWCNT**

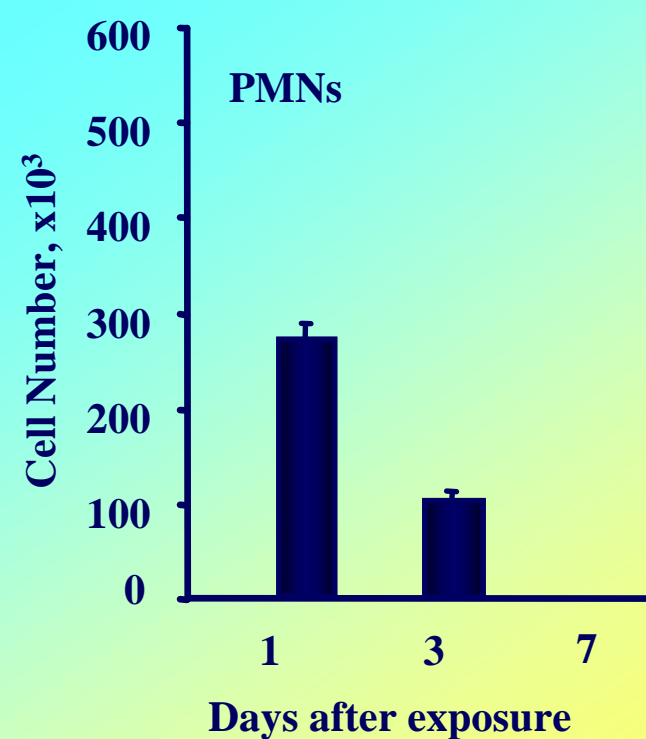
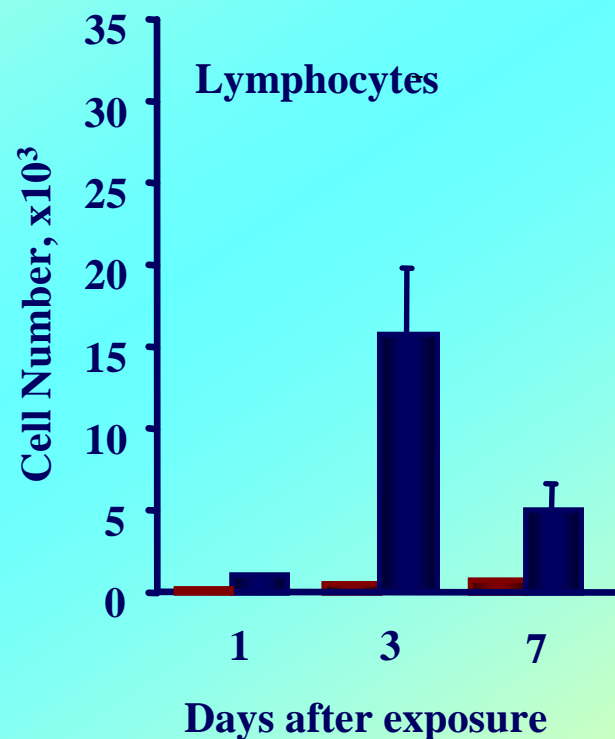
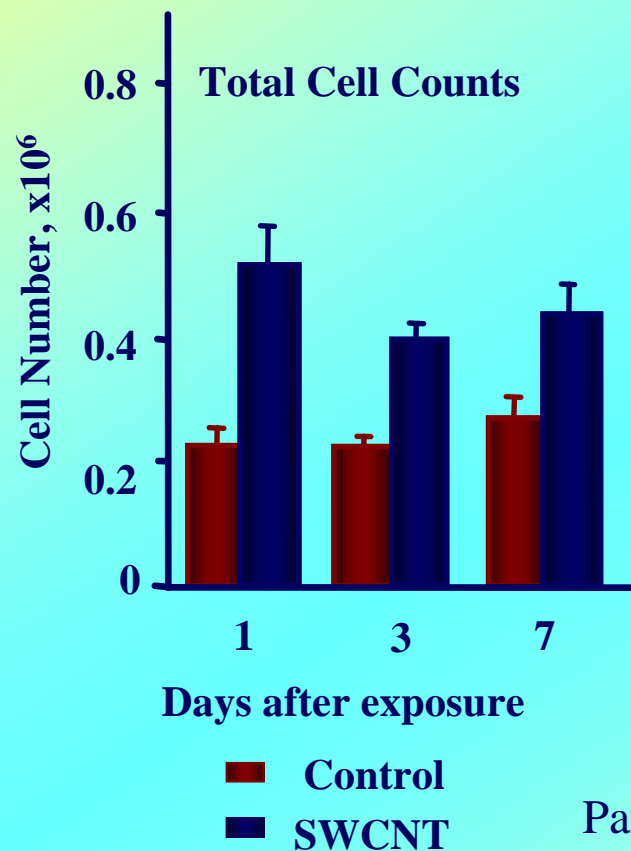


## Histopathology of lung of C57BL/6J mice after pharyngeal aspiration of partially purified SWCNT



Partially purified SWCNT (2.5 wt% of iron, 0.5 mg/kg b. w.).

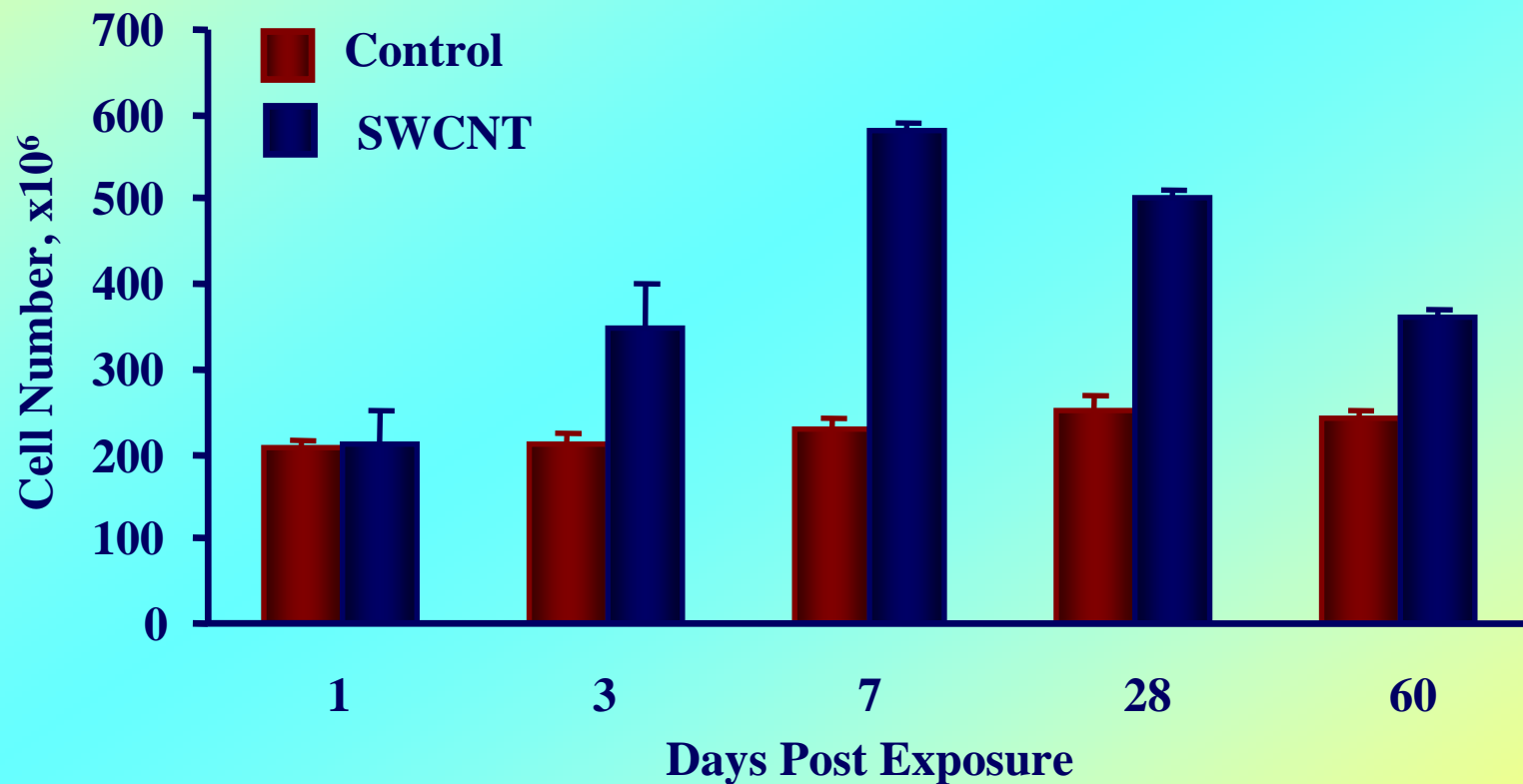
## Cell differential in BAL fluid of C57BL/6J mice after treatment with partially purified SWCNT.



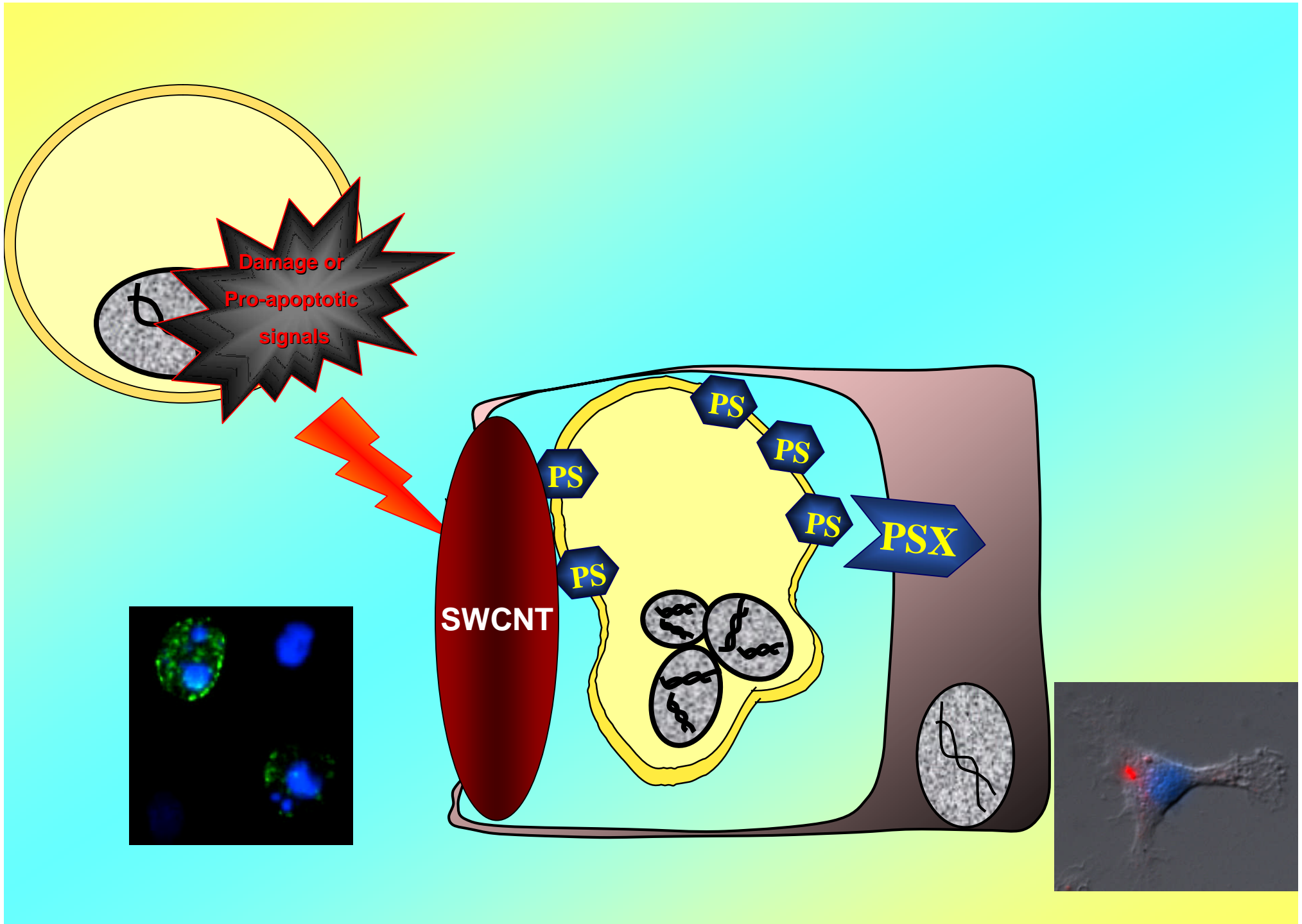
Partially purified SWCNT (2.5 wt% of iron, 0.5 mg/kg b.w.)



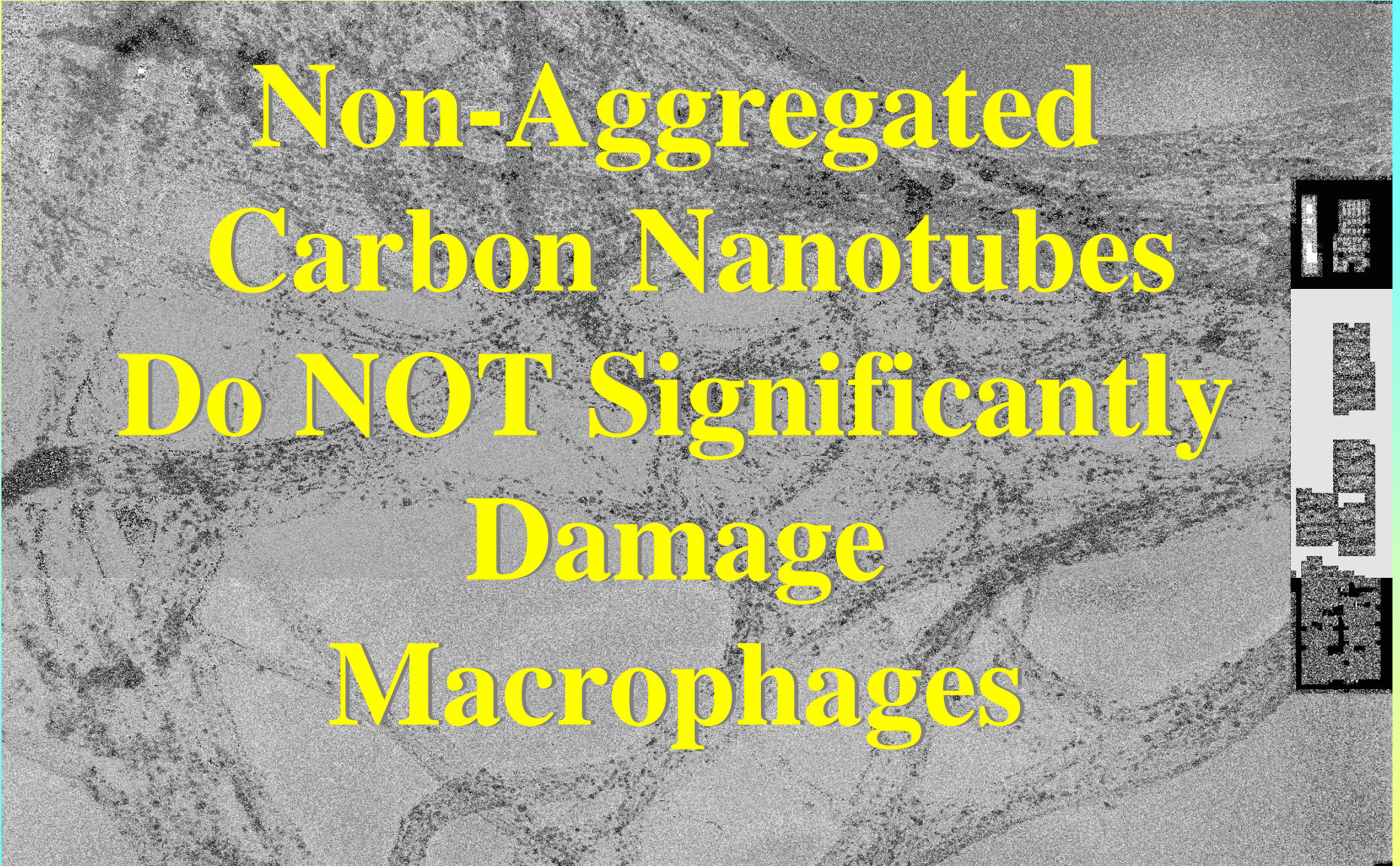
## Macrophages in BAL fluid of C57BL/6J mice after treatment with partially purified SWCNT.



Partially purified SWCNT (40 µg per mouse)





The background of the slide is a grayscale electron micrograph showing a dense network of carbon nanotubes. The nanotubes appear as thin, dark, wavy lines against a lighter, textured background. On the right side of the micrograph, there is a vertical scale bar with several rectangular segments of varying heights and widths, likely used for size calibration.

**Non-Aggregated  
Carbon Nanotubes  
Do NOT Significantly  
Damage  
Macrophages**



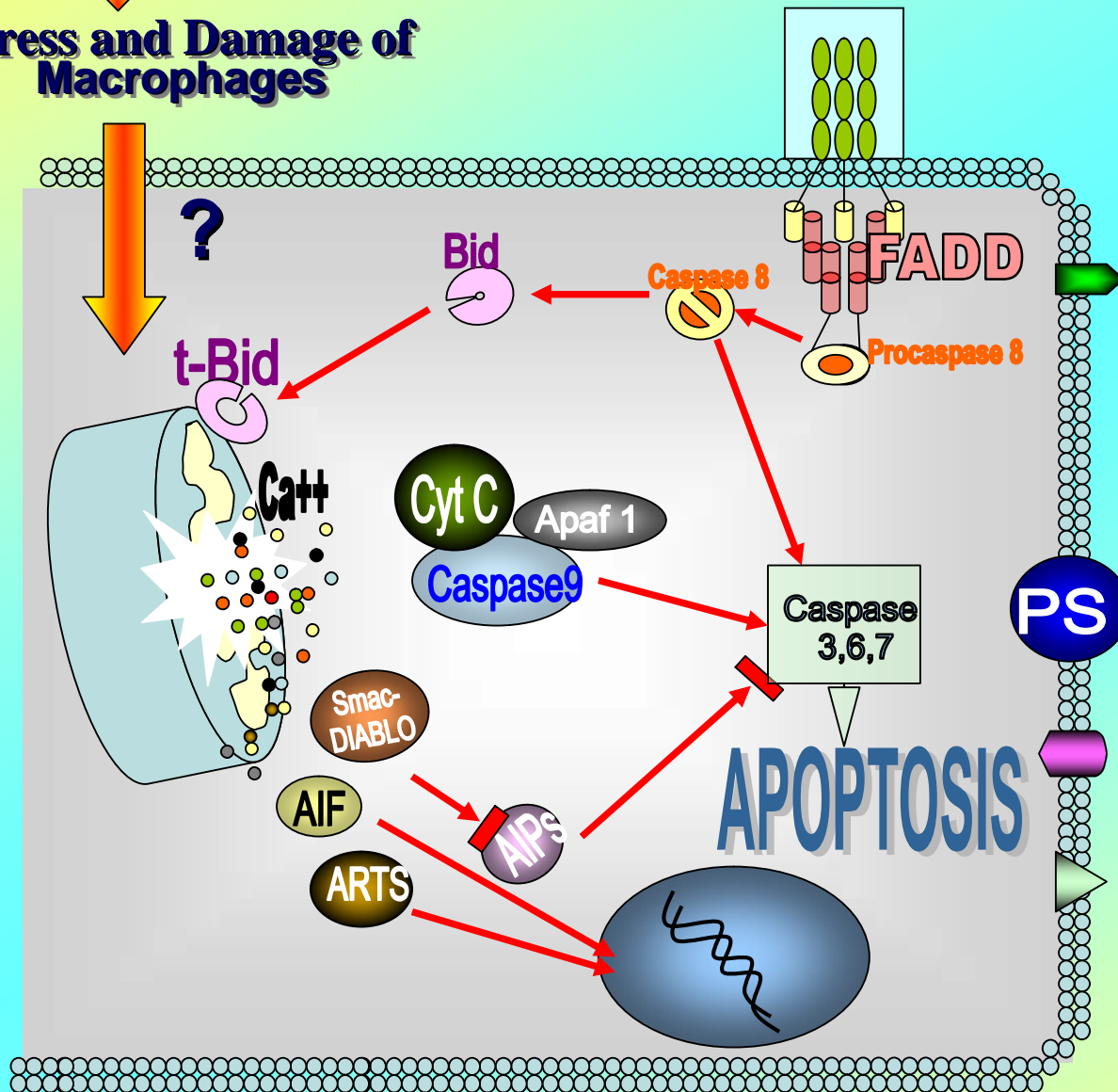
# ΑΠΟΠΤΟΣΙΣ

30 2:57 PM



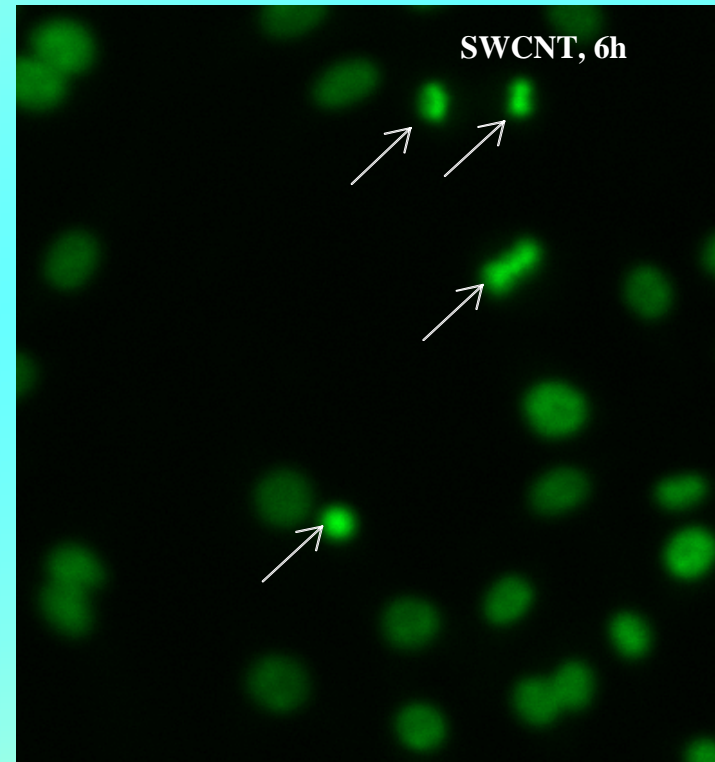
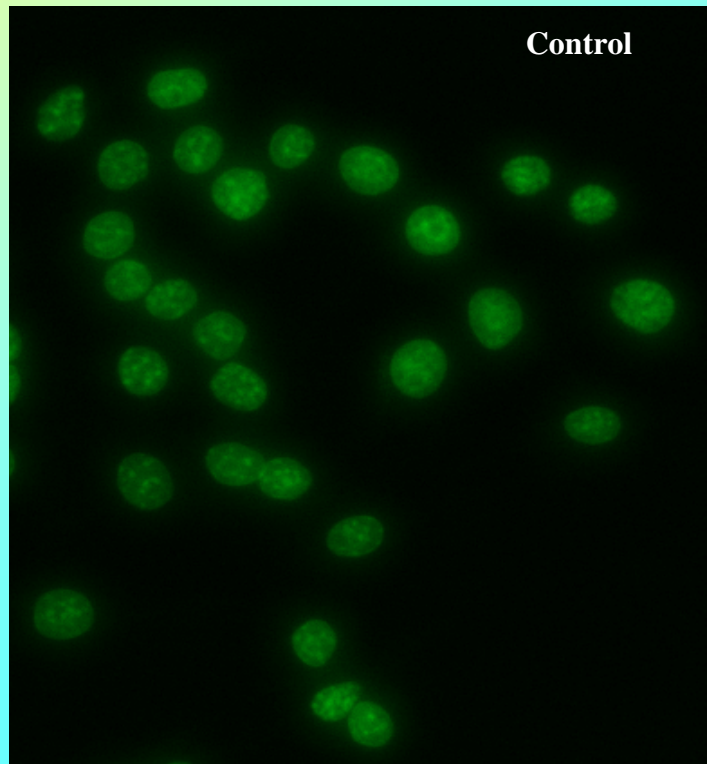
# Carbon Nanotubes

Stress and Damage of Macrophages



**Do  
Carbon  
Nanotubes  
Induce  
Apoptosis  
in  
Macrophages?**

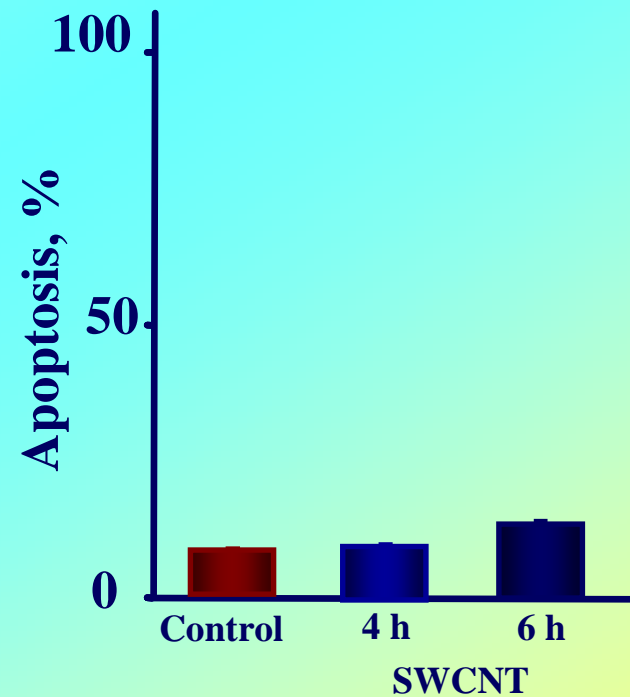
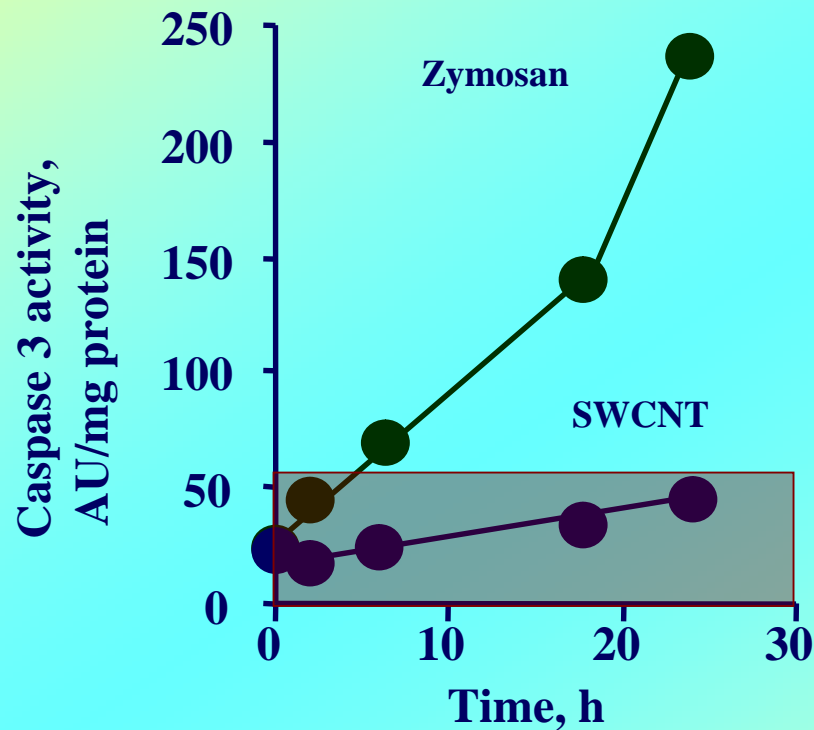
# Fluorescent micrographs of RAW 264.7 macrophages incubated in the presence of partially purified SWCNT.



Note nuclear condensation and fragmentation as revealed by Hoechst 33342 staining (shown for clarity in green pseudo-color).

Partially purified SWCNT (2.5 wt% of iron, 0.12 mg/ml).

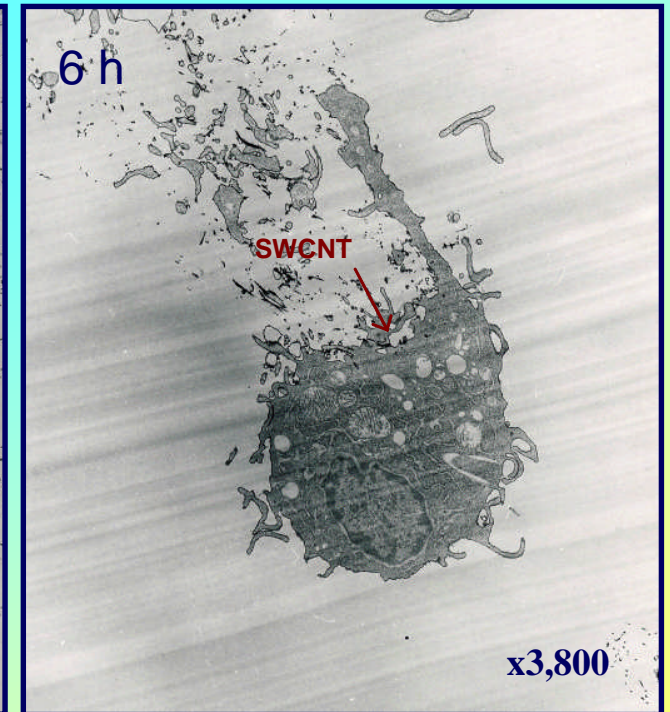
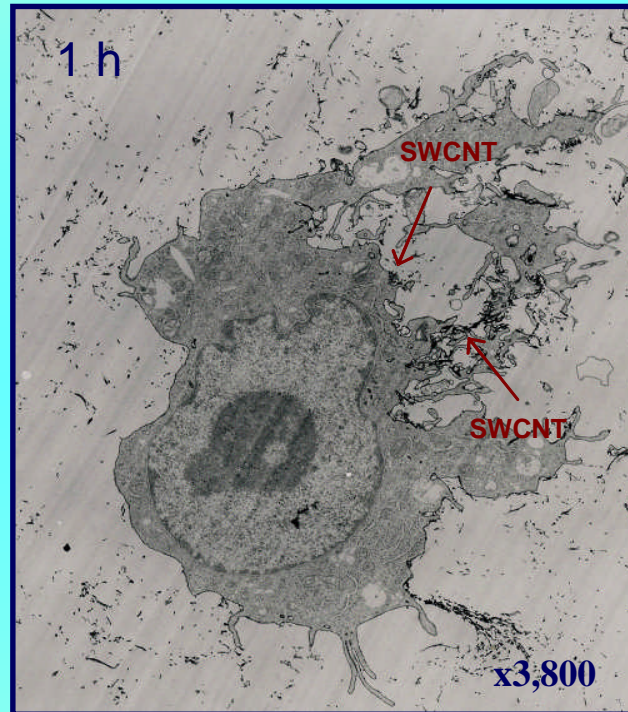
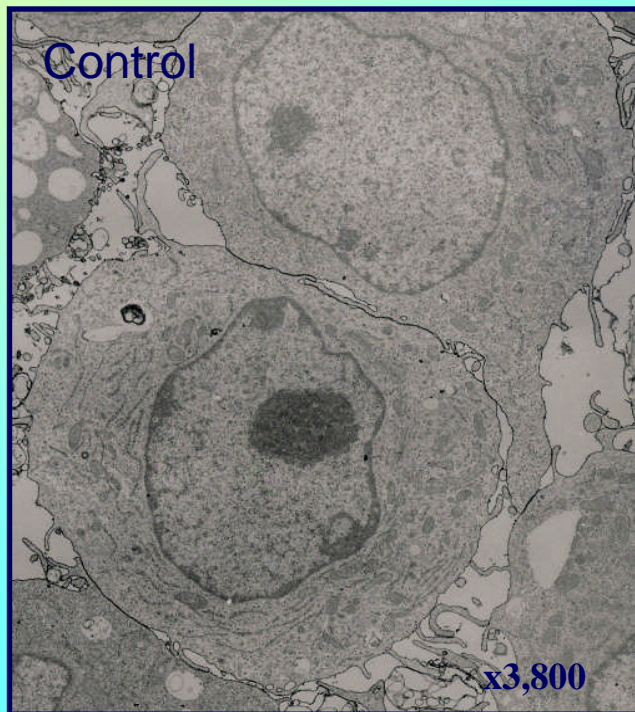
# Apoptosis induced by partially purified SWCNT in RAW 264.7 macrophages .



Partially purified SWCNT (2.5 wt% of iron, 0.1 mg/ml).



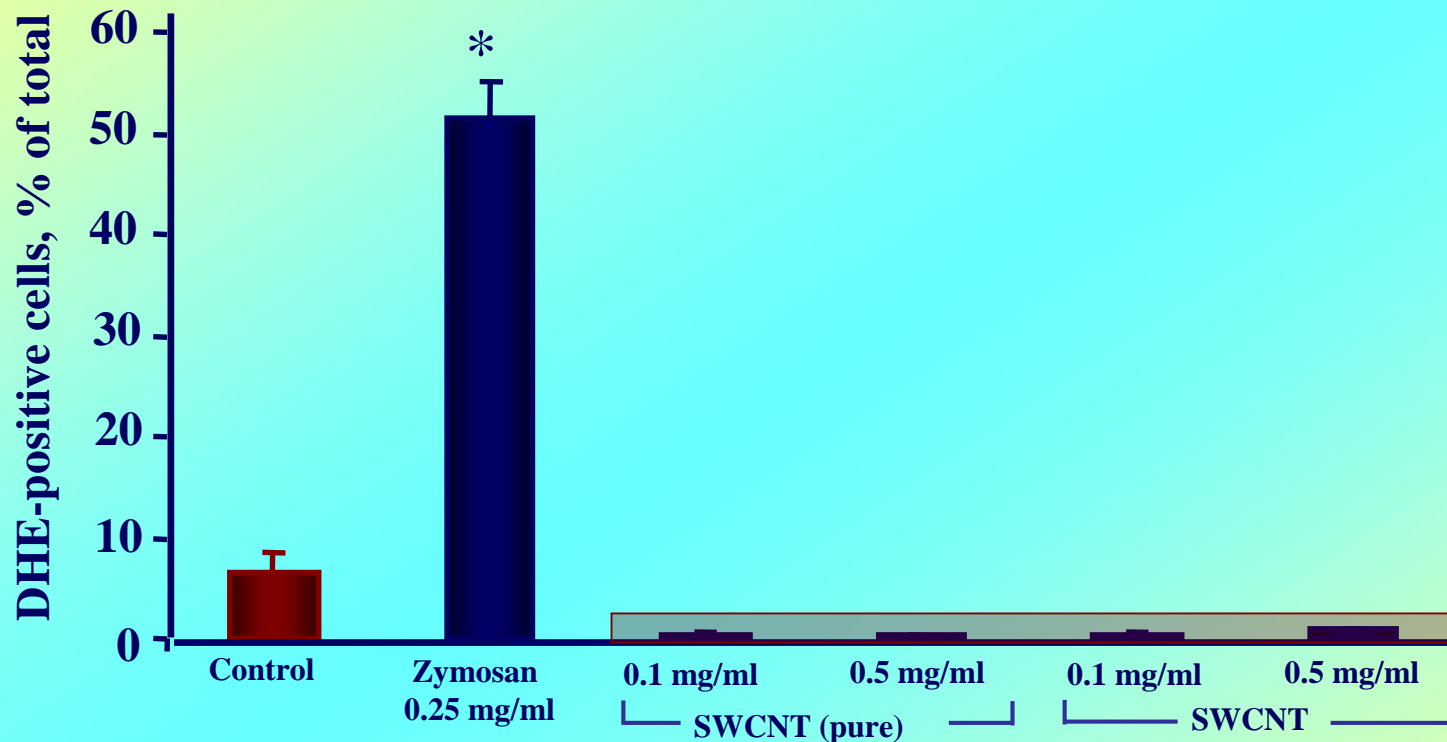
# Electron micrographs illustrating effects of partially purified SWCNT exposure on RAW 264.7 macrophages.



Partially purified SWCNT exposure (2.5 wt% of iron, 0.12 mg/ml)

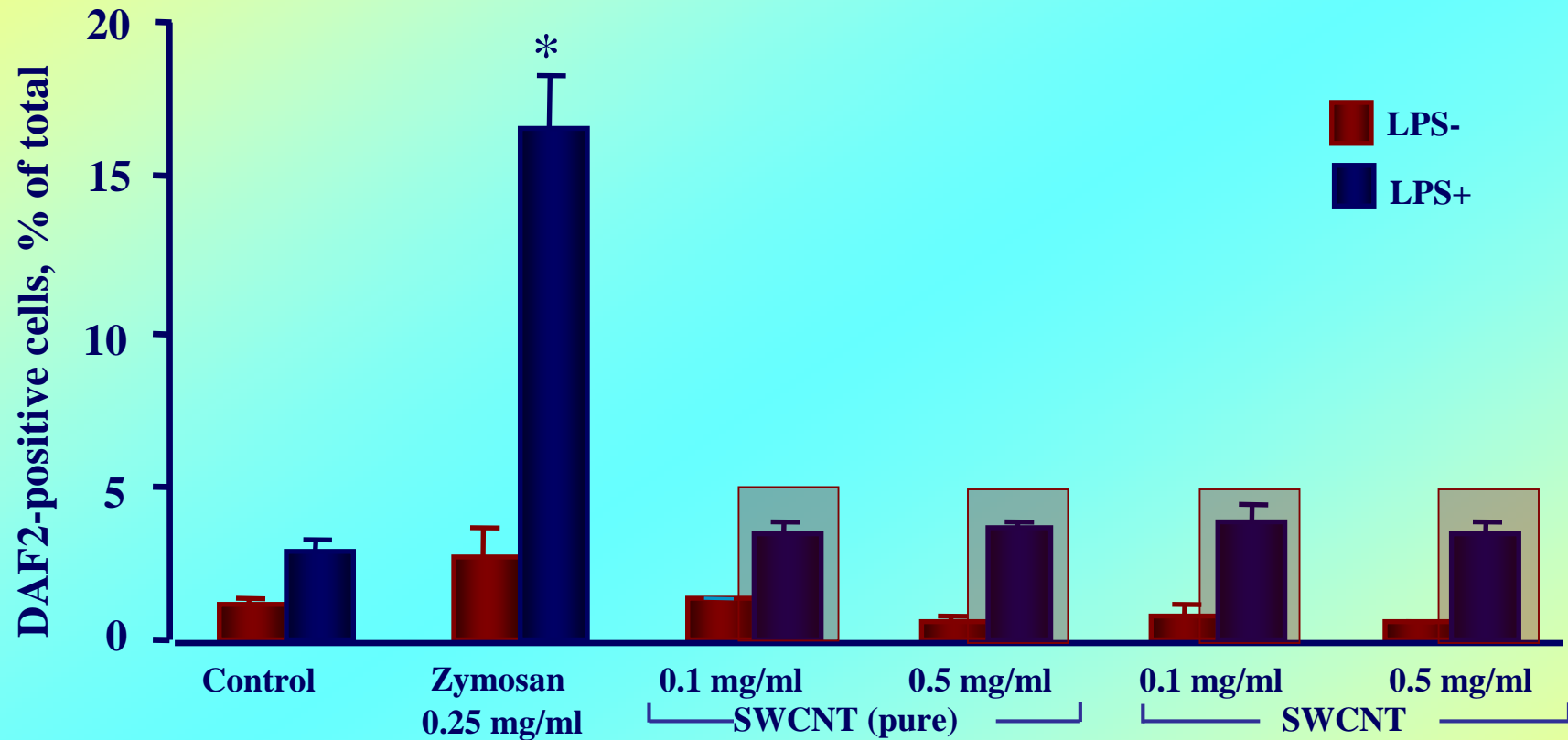


## DHE-positive RAW 264.7 macrophages: zymosan and SWCNT.

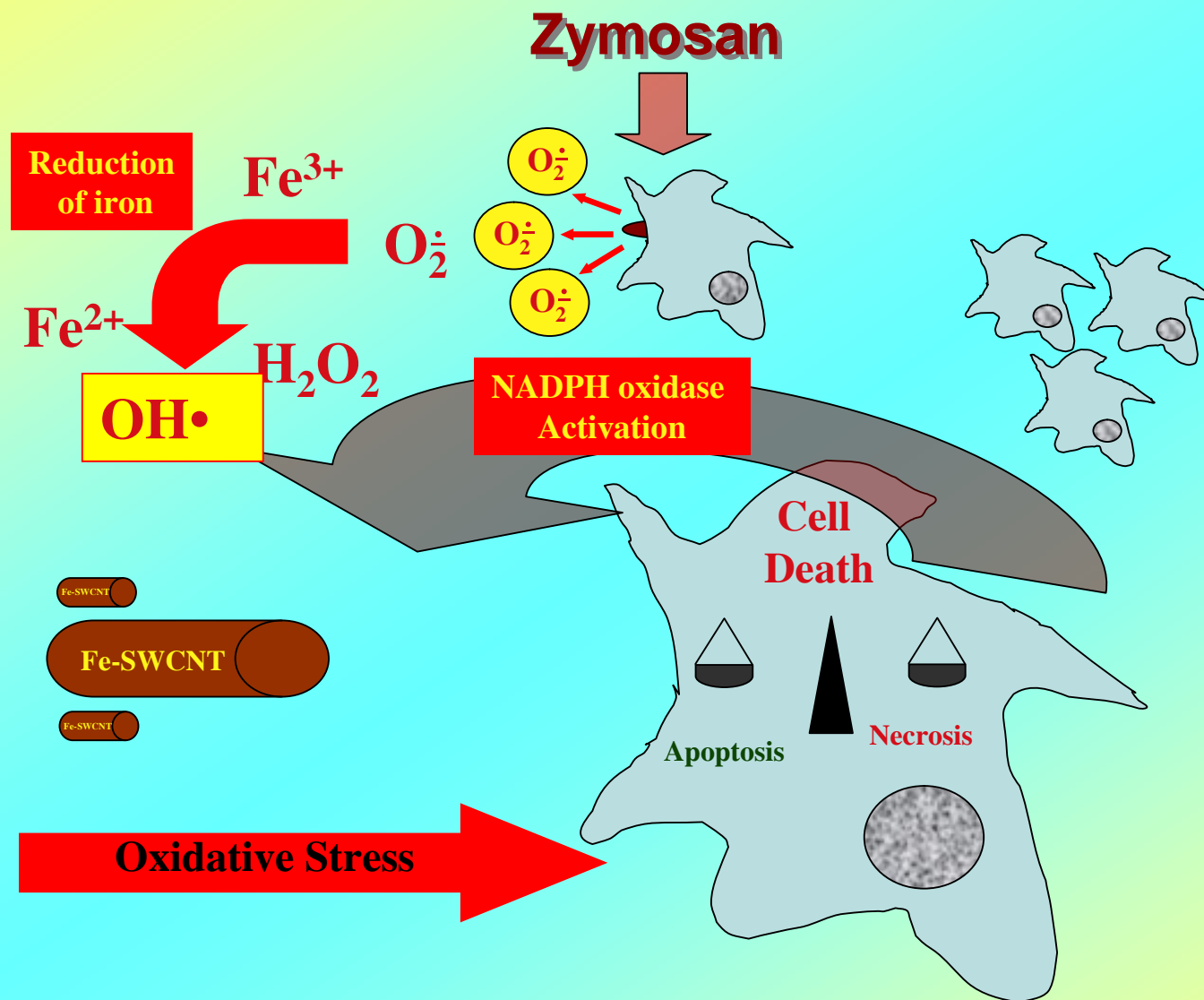


Macrophages ( $0.3 \times 10^6$ /well) were pre-incubated with DHE (10 mM for 10 min at 37°C). Then RAW 264.7 macrophages were stimulated by zymosan or SWCNT (for 30 min at 37°C). \* -  $p < 0.05$  vs. control cells

## DAF2-positive RAW 264.7 macrophages: zymosan and SWCNT.

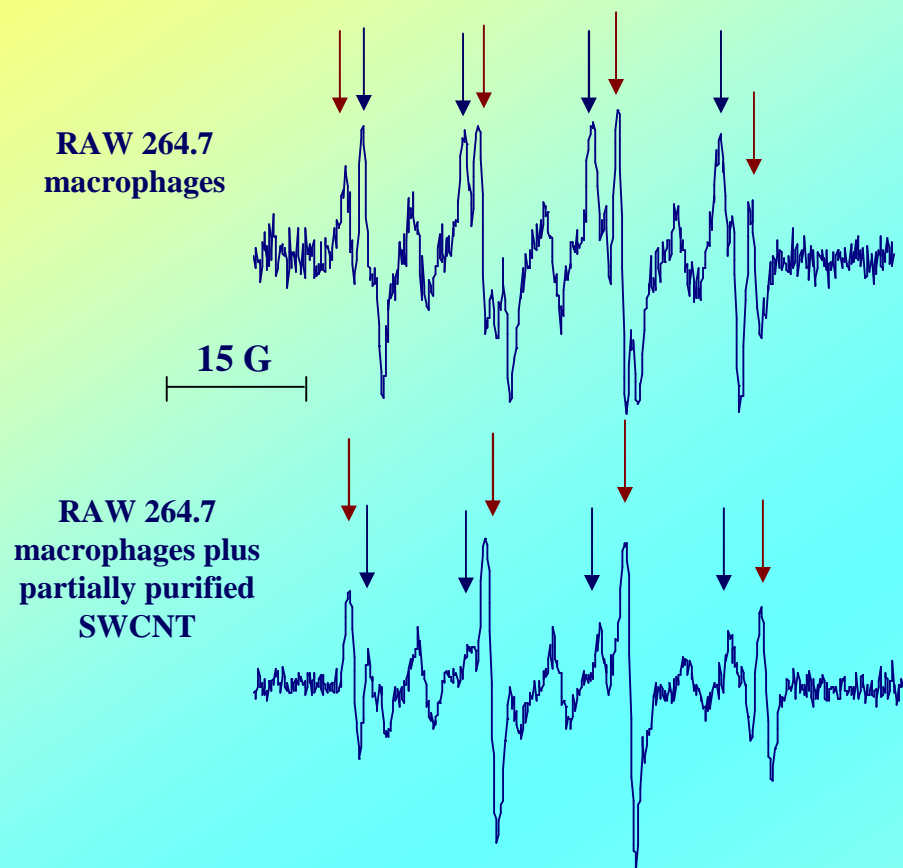


Naïve macrophages ( $0.3 \times 10^6$ /well) and macrophages stimulated by LPS (0.1 mg/ml for 6 h at 37°C) macrophages were pre-incubated with DAF-2DA (2 mM for 1 h at 37°C). Then RAW 264.7 macrophages were stimulated by zymosan or SWCNT (for 2h at 37°C). \* -  $p < 0.05$  vs. control cells



## Superoxide-DMPO Adduct

**EPR spectra of DMPO radical adducts generated during incubation of xanthine oxidase/xanthine and zymosan-stimulated RAW 264.7 macrophages by partially purified SWCNT in the absence or presence of partially purified SWCNT.**

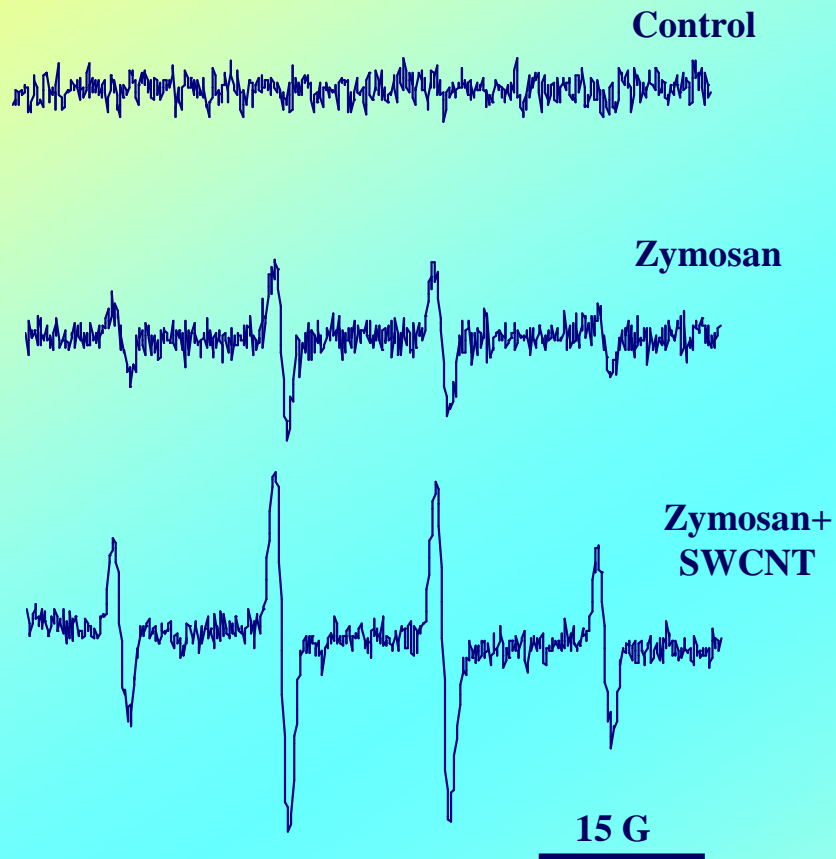


## Hydroxyl Radical-DMPO Adduct

Incubation system contained: xanthine oxidase (0.1U/mL), xanthine (1 mM), zymosan (2.5 mg/mL)-stimulated RAW264.7 macrophages ( $20 \times 10^6$  cells/ml) in PBS (pH 7.4) plus 100 mM DMPO; partially purified SWCNT (2.5 wt% of iron, 0.12 mg/ml); EPR conditions: microwave power, 20 mW; modulation amplitude, 1.0 G; time constant, 1.3 sec; conversion time, 0.6 sec.



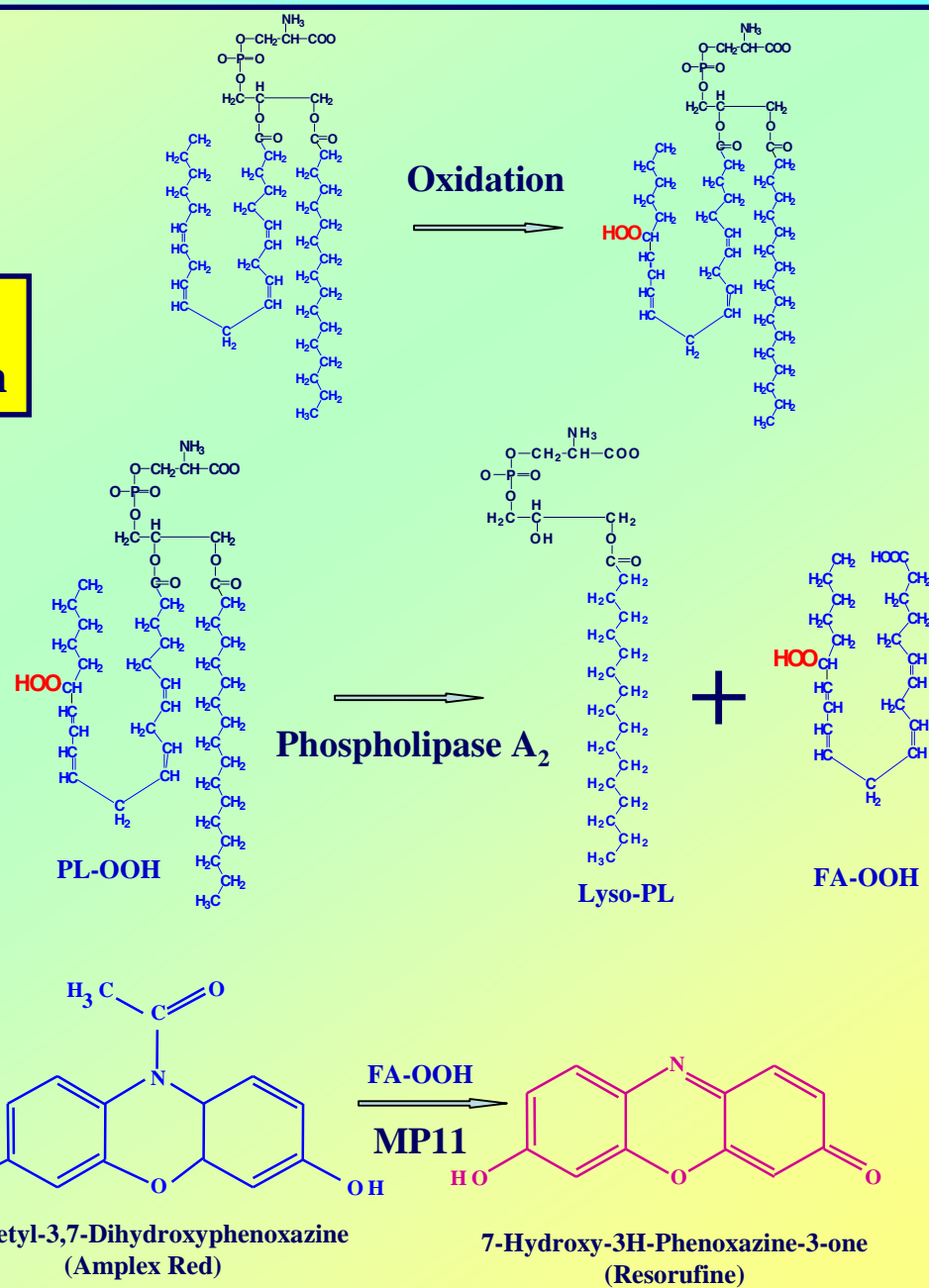
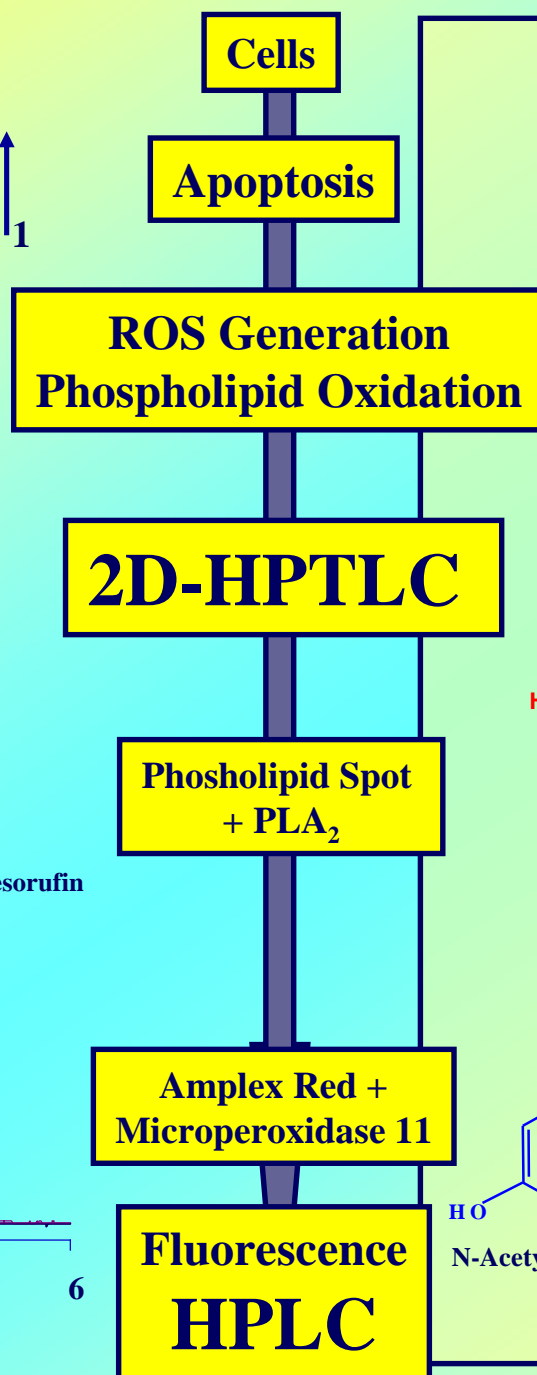
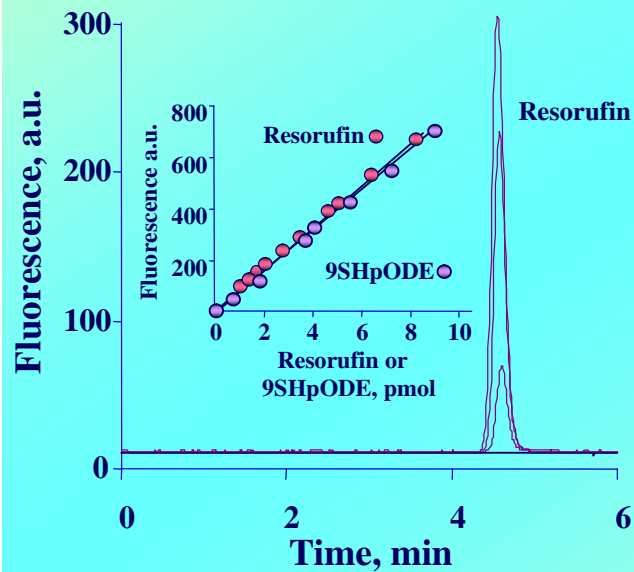
## Hydroxyl Radical-DMPO Adduct



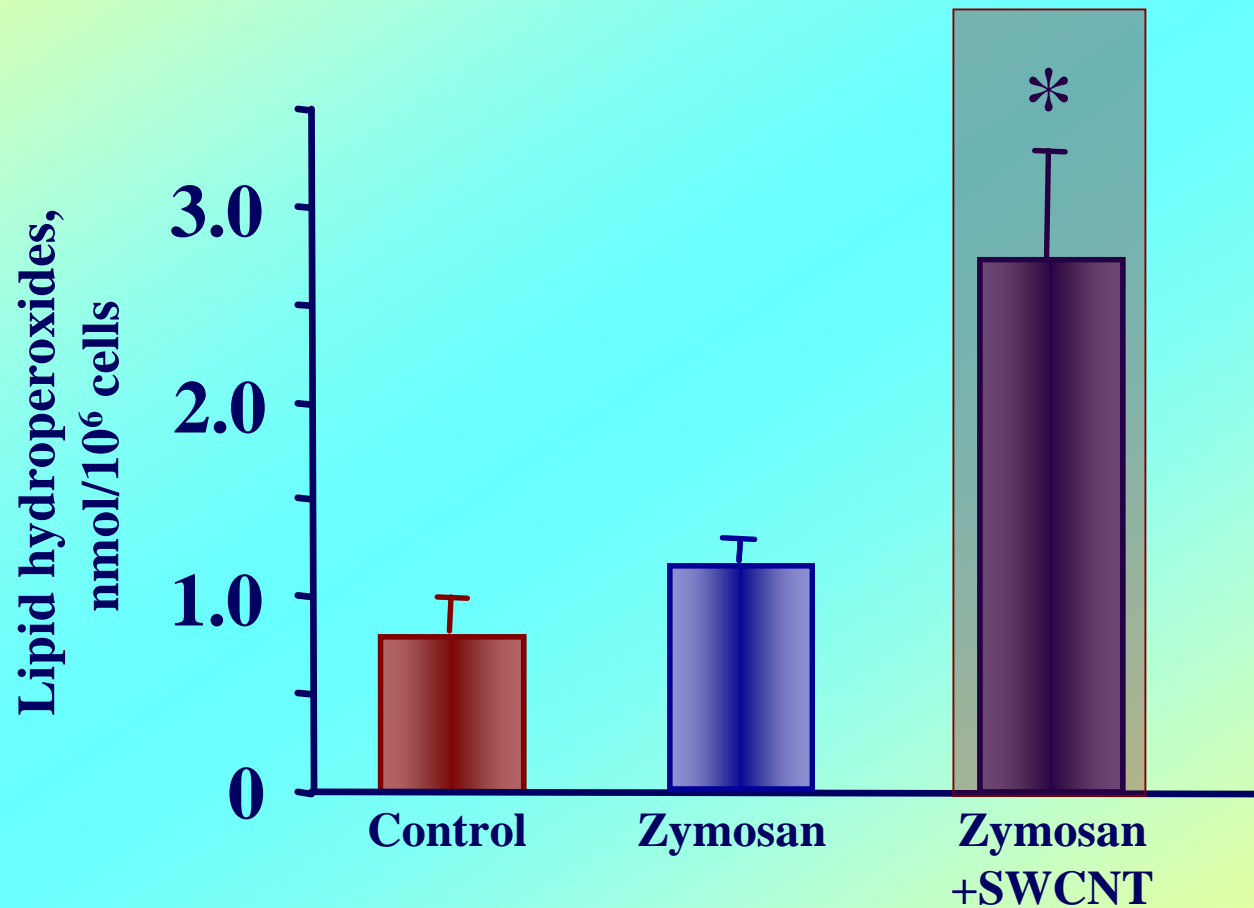
**ESR spectra of  
DMPO adducts of  
free radicals formed  
by partially purified  
SWCNT in the  
presence of RAW  
264.7 macrophages**

1

2

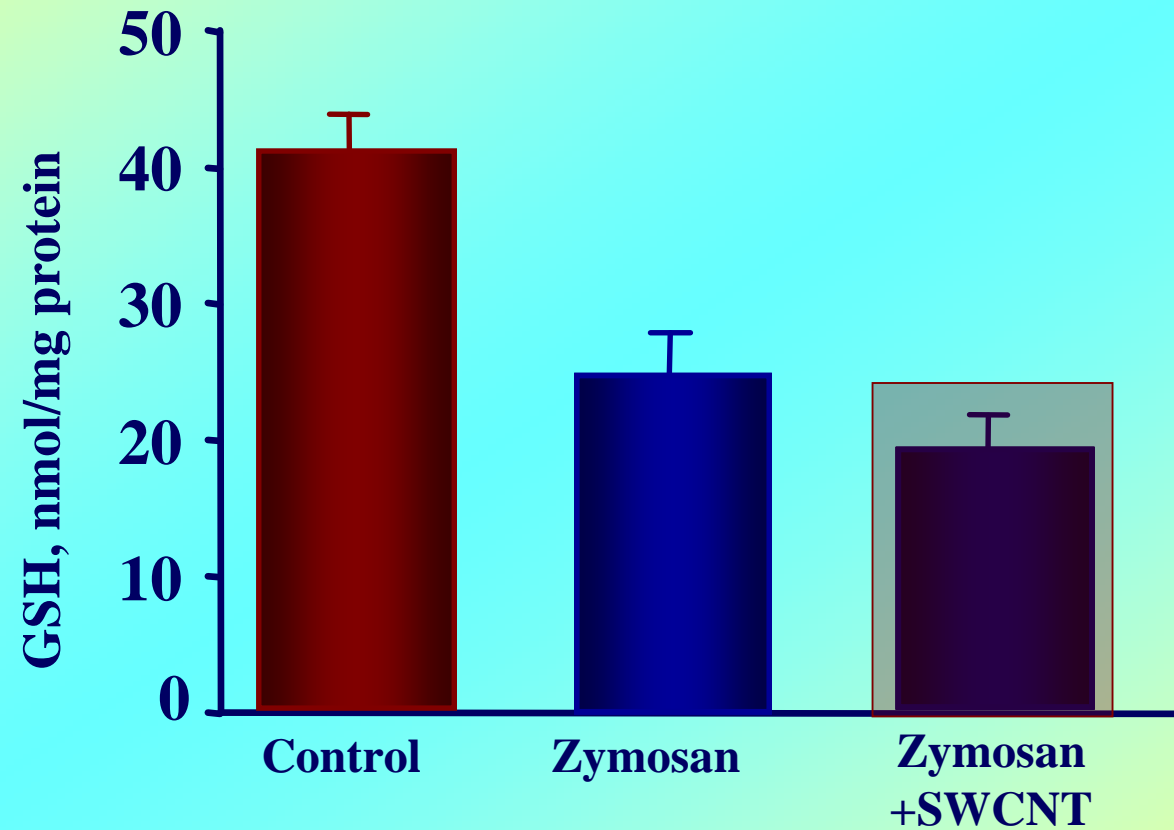


## Partially purified SWCNT induce lipid peroxidation in RAW 264.7 macrophages .



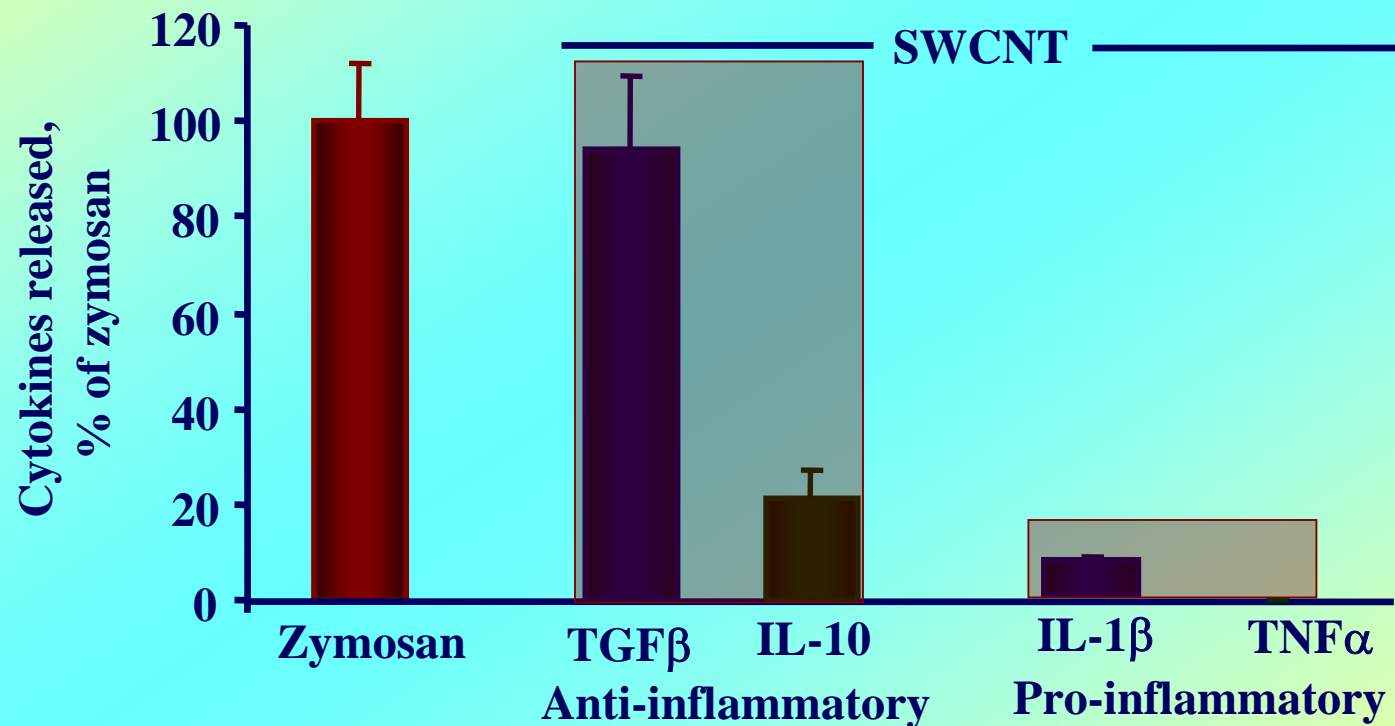
Stimulation of macrophages with zymosan (0.25 mg/ml) cause a slight increase in lipid peroxidation further enhanced by partially purified iron-containing SCWNT (0.1 mg/ml). Lipid peroxidation was assessed by our newly developed fluorescence HPLC-based protocol for lipid hydroperoxides

## Effect of partially purified SWCNT exposure on GSH content in RAW 264.7 macrophages.





## Production of cytokines by zymosan-stimulated RAW 264.7 macrophages and SWCNT treated RAW 264.7 macrophages.

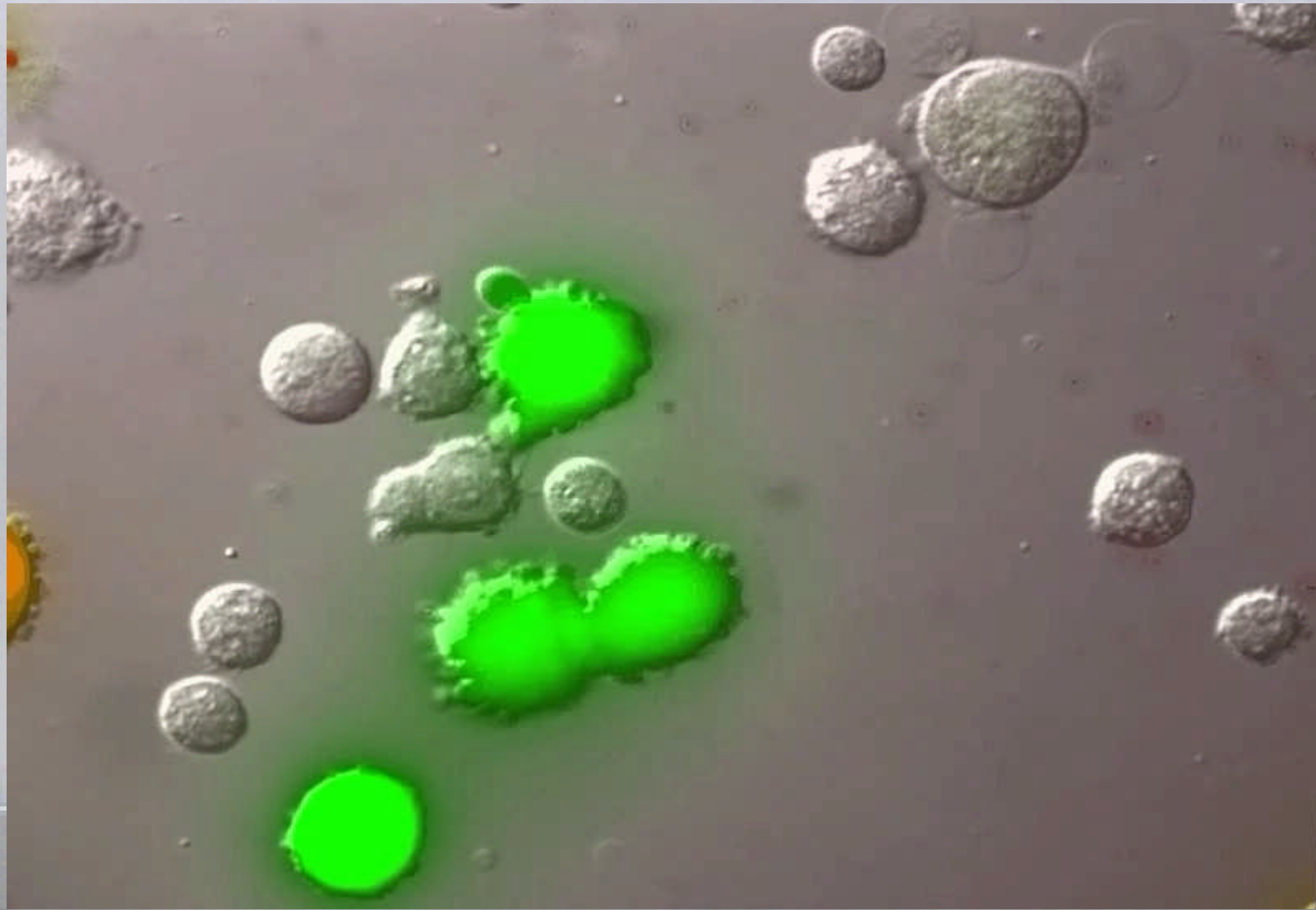


Zymosan- 0.25 mg/ml; SWCNT 0.1 mg/ml.



# Clearance, Phagocytosis

# **Recognition and digestion of Cells**

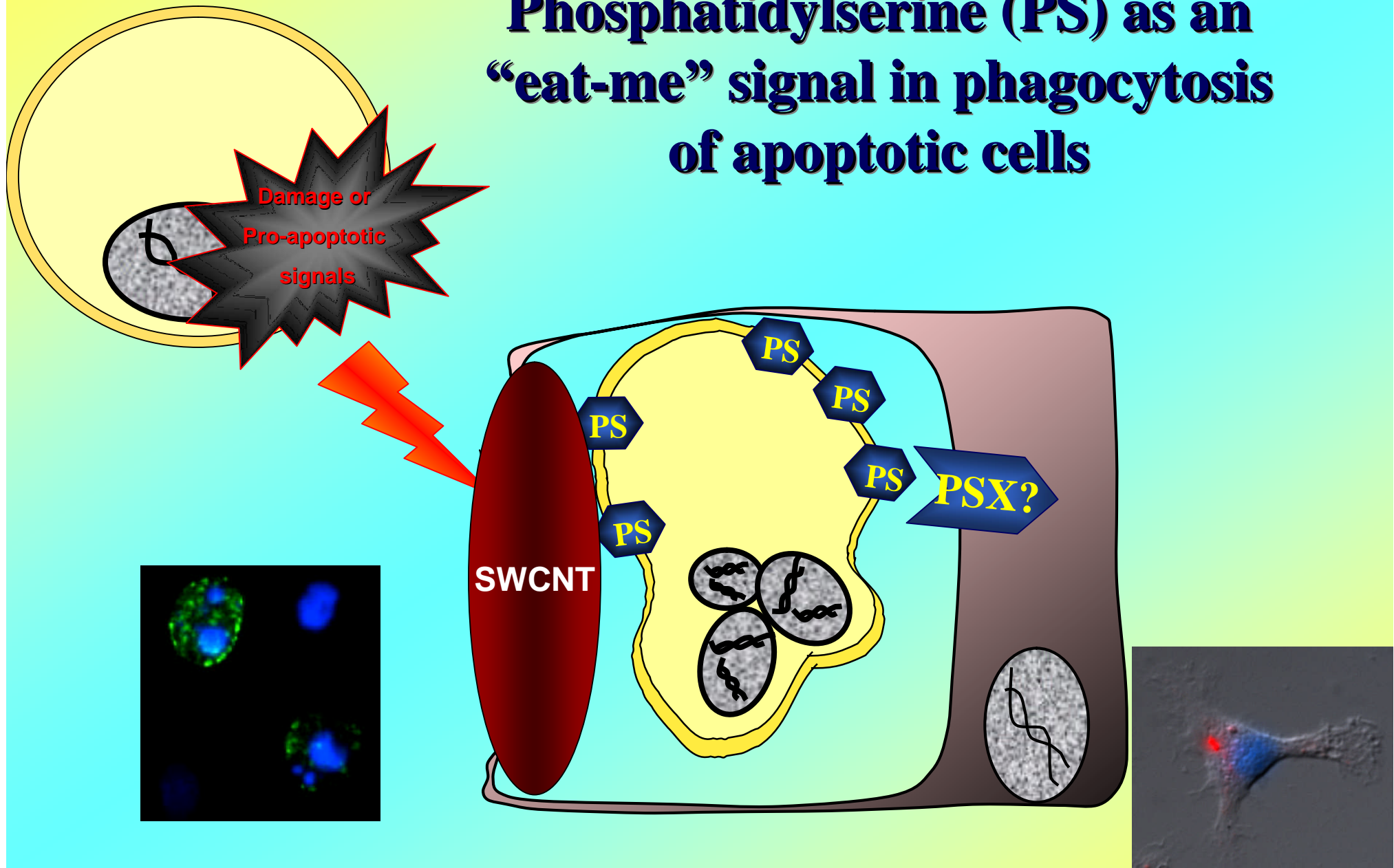


**B16 melanoma cells  
(green)  
and dendritic cells**

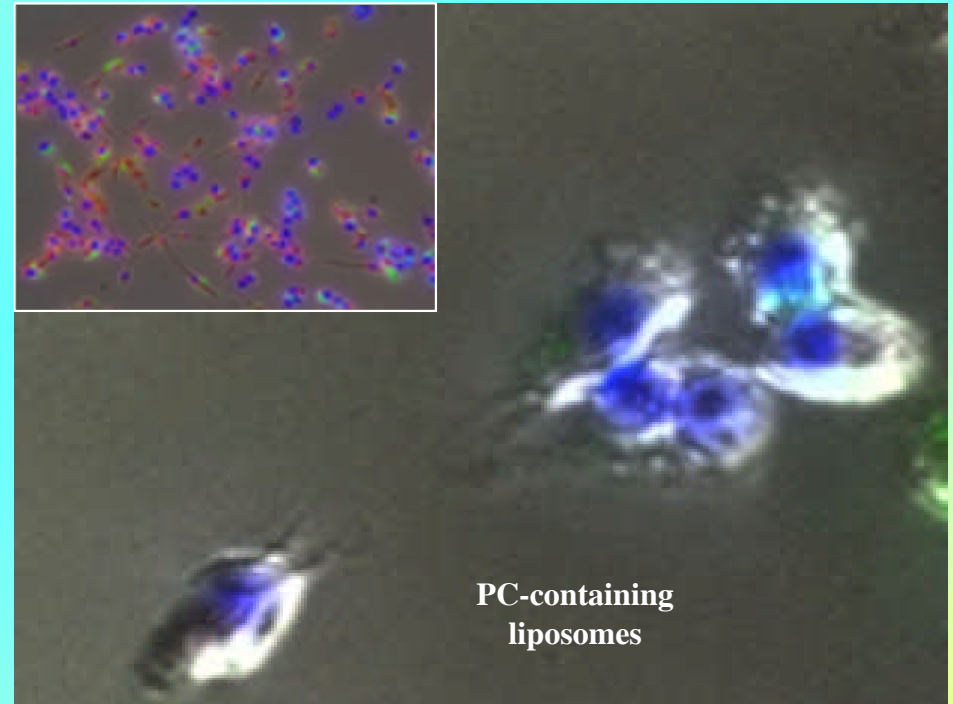
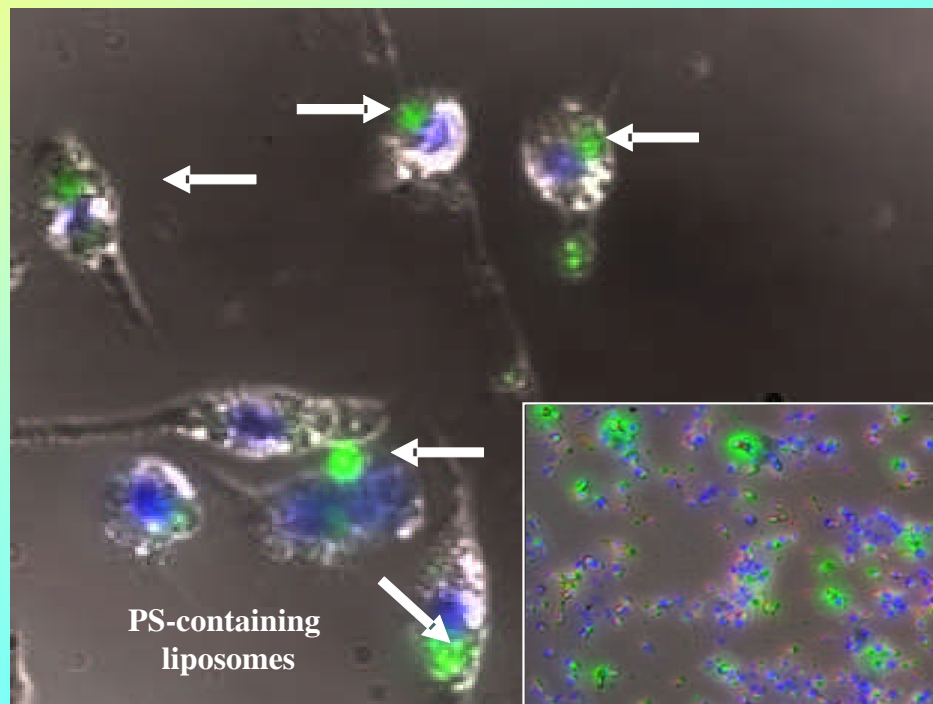


**Can Macrophages  
Be Forced  
to Recognize and  
Digest  
Carbon  
Nanotubes**

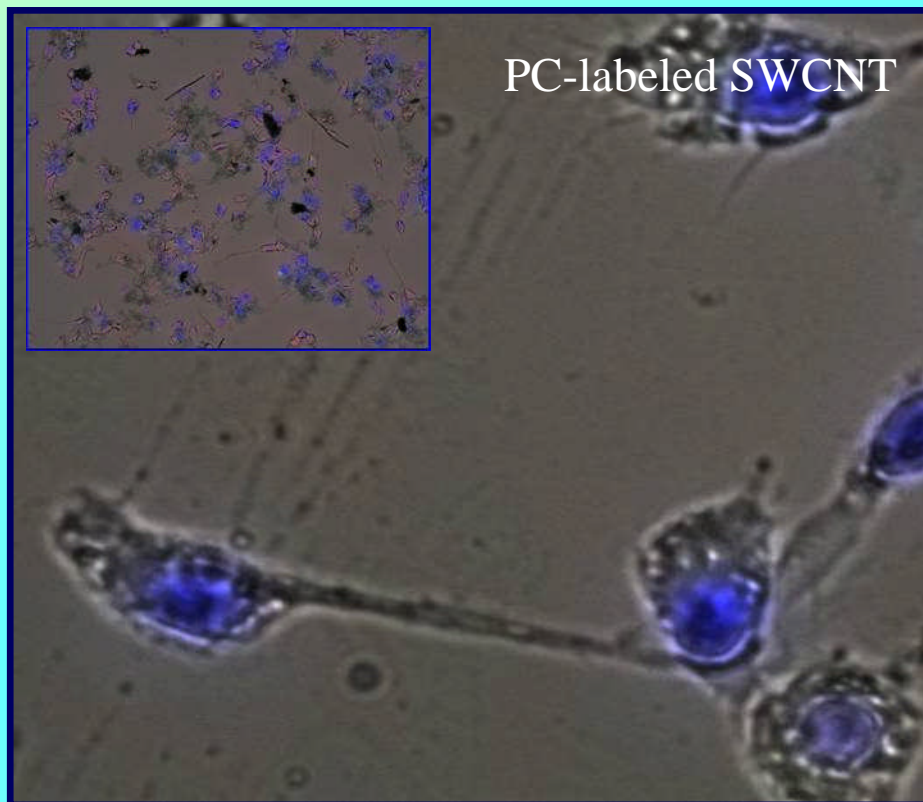
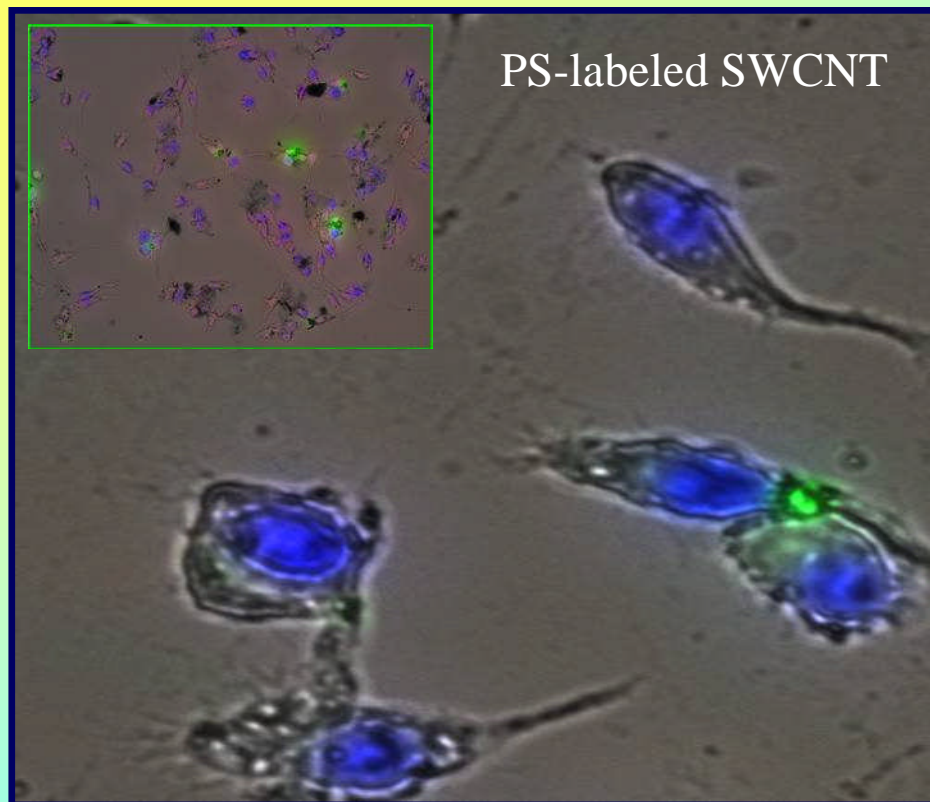
# Phosphatidylserine (PS) as an “eat-me” signal in phagocytosis of apoptotic cells



## **RAW 264.7 macrophages effectively phagocytose PS-containing liposomes but not PC-containing liposomes.**



RAW 264.7 macrophages ( $10^5$  cells/ml) were incubated for 6 h with liposomes (0.33 mM) composed of a mixture of PC:PS (with fluorescently labeled PS) or PC (with fluorescently labeled PC). After incubation, macrophages were fluorescently labeled with Hoechst 3343 (nuclei, blue fluorescence), and Cell Tracker Orange (cytosol, red fluorescence). Liposomes fluorescently labeled with with NBD-phospholipids (green fluorescence) PS-containing liposomes were prepared by sonication of a mixture of PC:PS 1:1 with the addition of 10 mol% of NBD-PS). PC-containing liposomes were prepared by sonication of a mixture of PC with the addition of 10 mol% of NBD-PC.



**RAW 264.7 macrophages effectively phagocytose PS-labeled SWCNT but not PC labeled SWCNT.**

RAW264.7 macrophages ( $10^5$  cells/ml) were incubated for 6h with fluorescently labeled SWCNT (0.1 mg/ml).





# Thanks To My Collaborators:

## Kagan's Lab:

A. Arroyo  
N. Belikova  
G. Borisenko  
J. Jiang  
K. Kawai  
V. Kini  
S.-X. Liu  
T. Matsuura  
A. Osipov  
A. Potapovich  
B. Serinkan  
V. Tyurin  
Y. Tyurina  
Q. Zhao

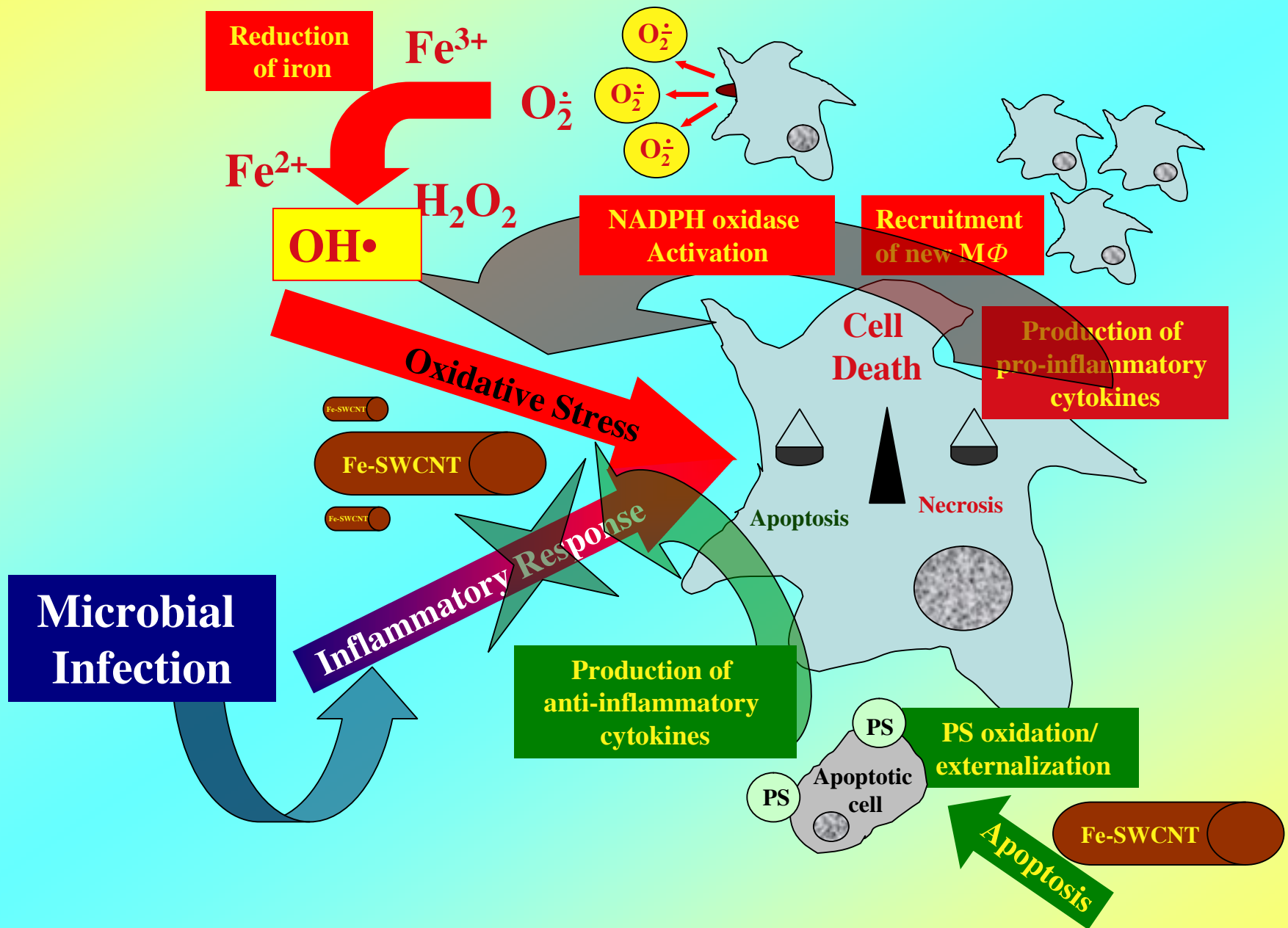
## NIOSH:

A. Shvedova (Morgantown)  
V. Castranova (Morgantown)  
R. Mercer (Morgantown)  
E. Kisin (Morgantown)  
A. Maynard (Cincinnati)



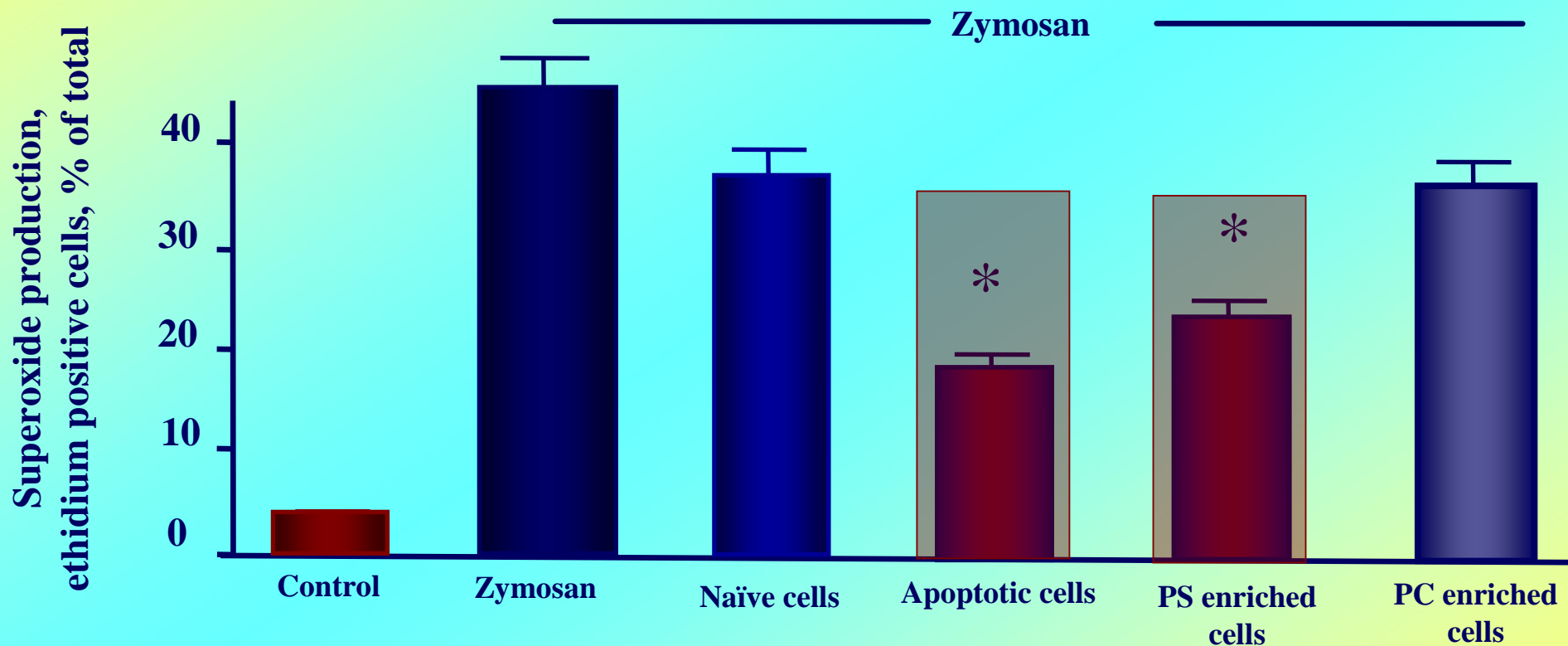






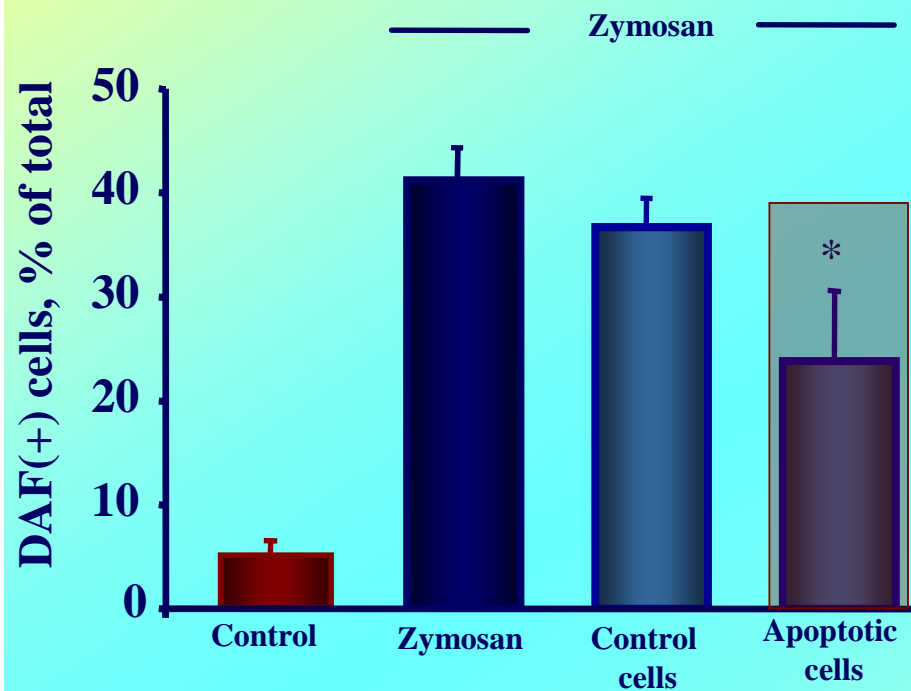


## Effect of apoptotic cells and PS on superoxide generation in zymosan-stimulated RAW 264.7 macrophages.

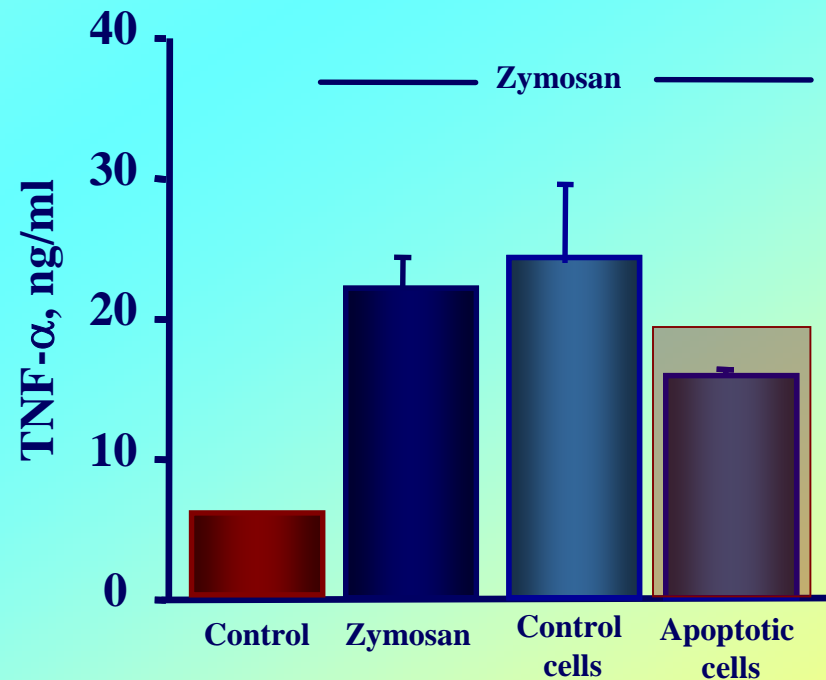


PS - 150 nmol/10<sup>6</sup> cells (30 min at 37°C); PC - 150 nmol/10<sup>6</sup> cells (30 min at 37°C); Zymosan - 0.25 mg/ml (1h at 37°C); DHE - 10 µM.  
Apoptosis in Jurkat cells was induced by anti-FAS (250ng/10<sup>6</sup> cells, 4h at 37°C.)

**Effect of apoptotic cells on  
formation of NO• in LPS-induced  
zymosan-stimulated RAW 264.7  
macrophages.**

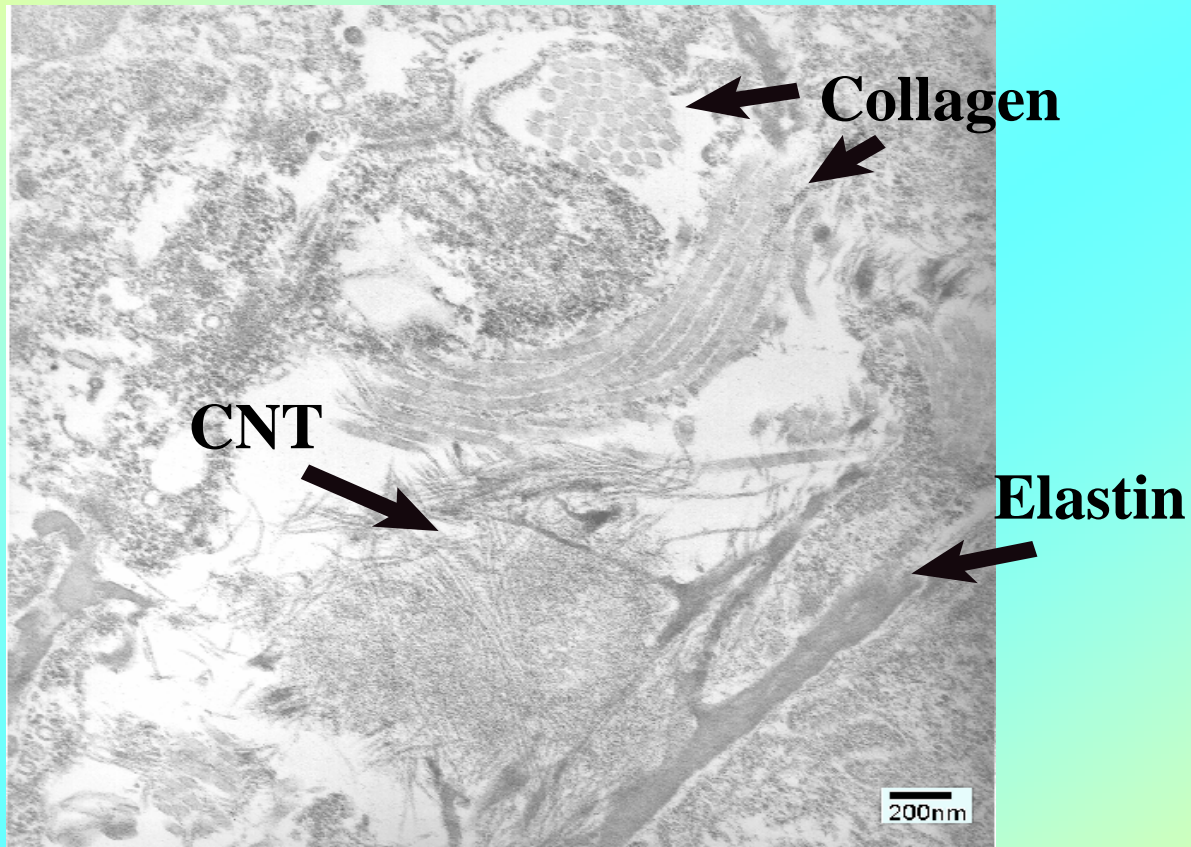


**Production of TNF $\alpha$  by  
zymosan-stimulated  
(0.25mg/ml) RAW 264.7  
macrophages.**



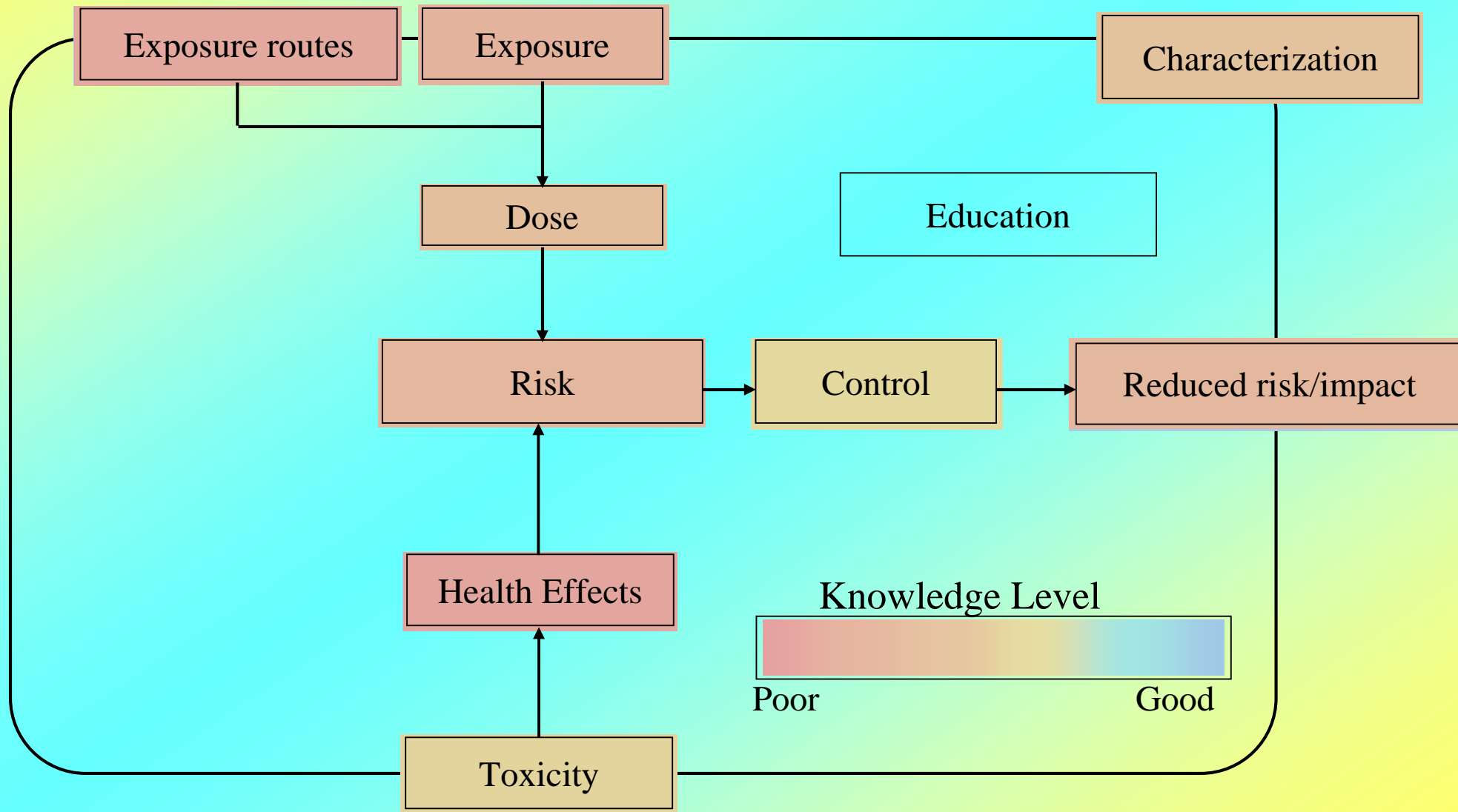
Zymosan - 0.25 mg/ml (1h at 37°C). Apoptosis in Jurkat cells was induced by anti-FAS (250ng/10<sup>6</sup> cells, 4h at 37°C.)

# Carbon Nanotubes in Interstitial Space



**TEM of carbon nanotubes in interstitial space. Micrograph shows carbon nanotubes intermixed with normal connective tissue matrix of the lungs.**

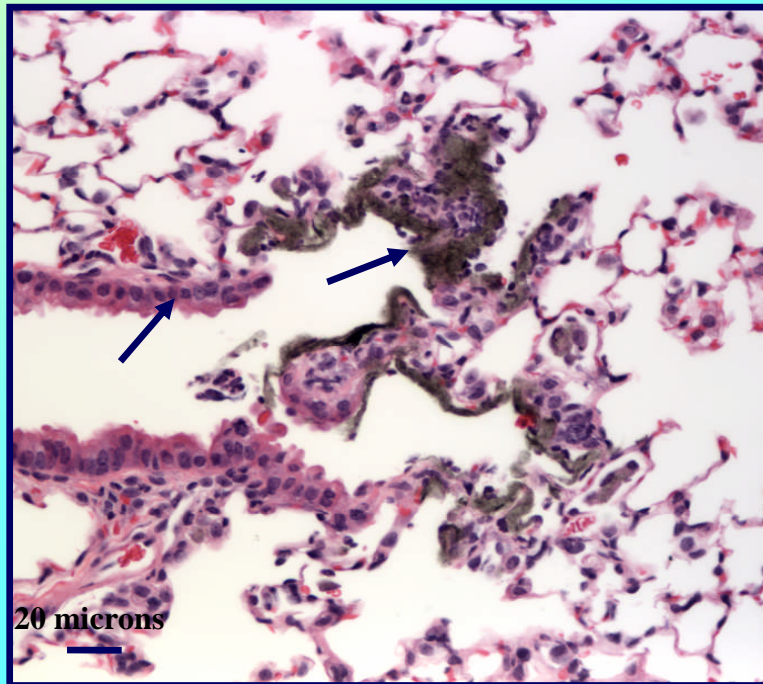
# Addressing occupational impact



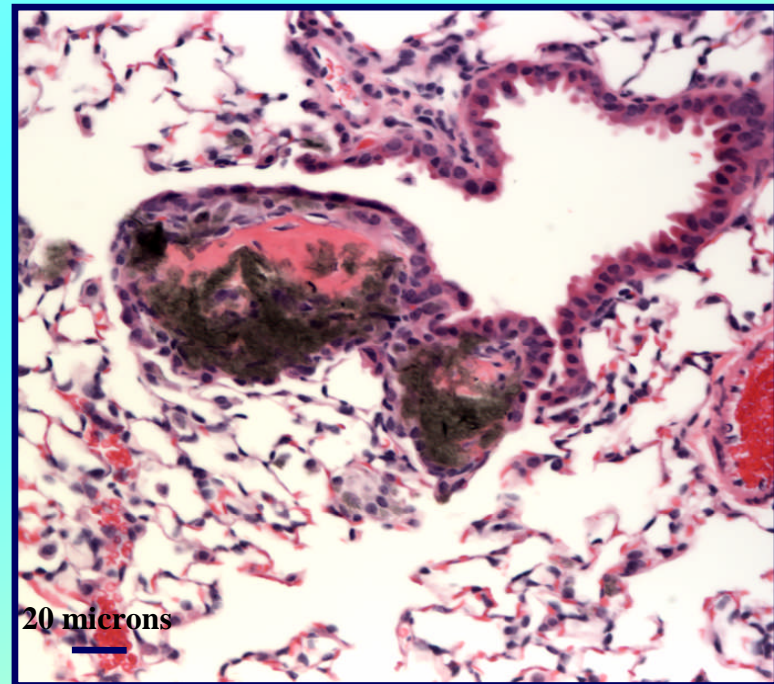


# Pharyngeal Aspiration of CNT in Mice Caused:

- Rapid development of granulomatous bronchointerstitial pneumonia
- Inflammation evolving with time from neutrophilic to granulomatous
- Morphologic alterations localized to the interstitial of the bronchiolar walls, alveolar ducts and adjacent alveoli
- Hypocellular eosinophilic material consistent with fibrous connective tissue observed within granulomas

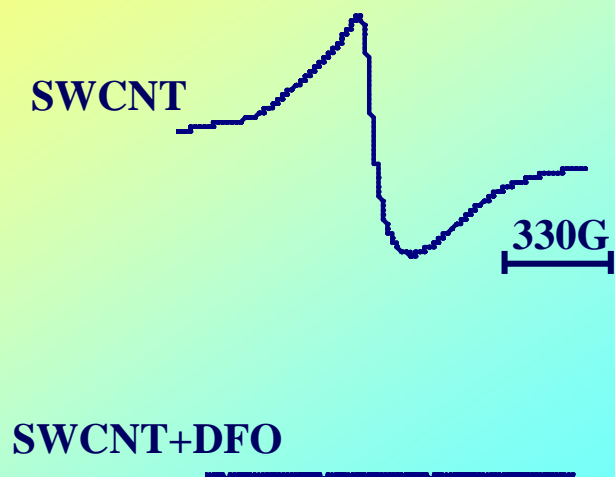


**1 day post exposure 40  
µg/mouse CNT**

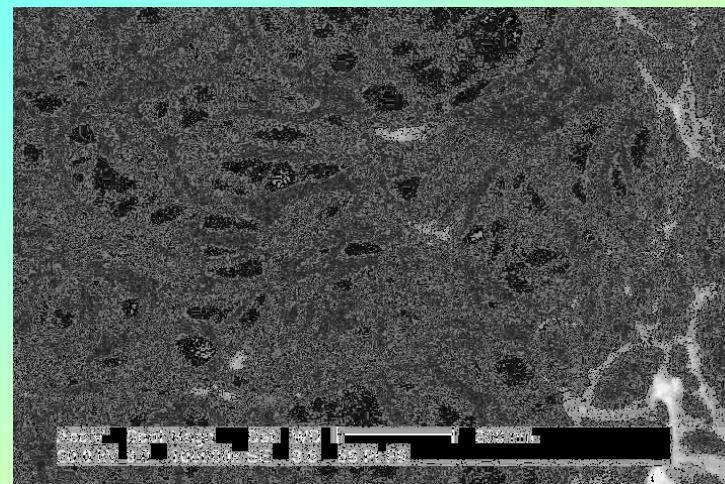
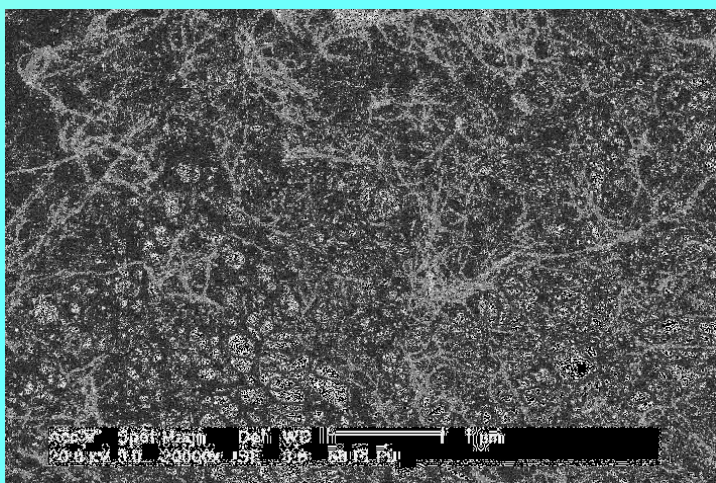


**28 days post exposure 40  
µg/mouse CNT**

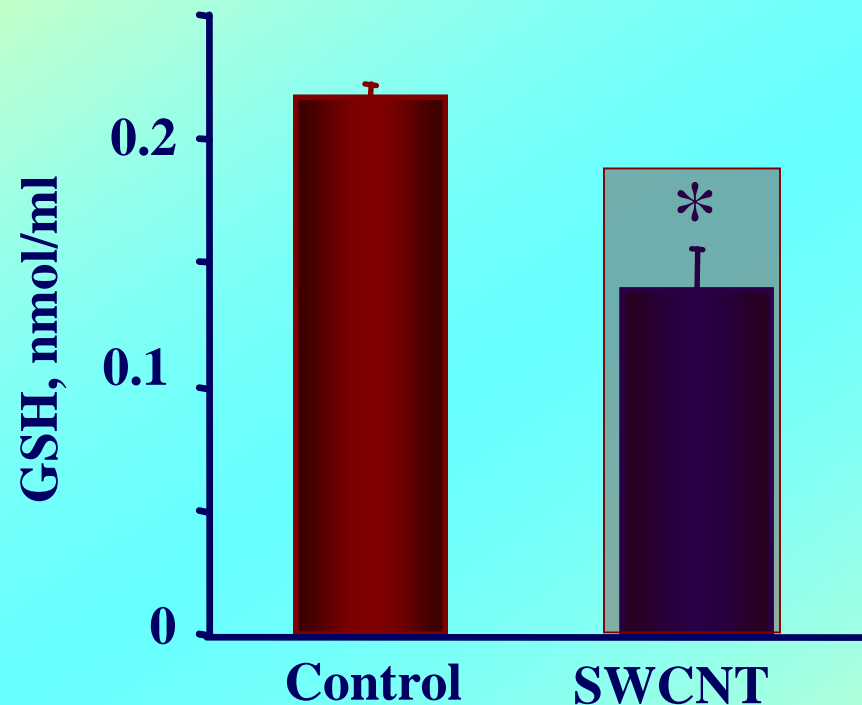
**EPR spectra of partially-purified SWCNT (0.5 mg/ml, 2.5wt% iron) manufactured by high-pressure CO conversion (HiPco™) technology as compared to purified SWCNT additionally treated with an iron chelator, deferoxamine (DFO).**



Note that partially-purified SWCNT displayed a broad signal with g value 2.0 and half-width of 640G, the signal was not detectable in purified DFO-treated SWCNT.



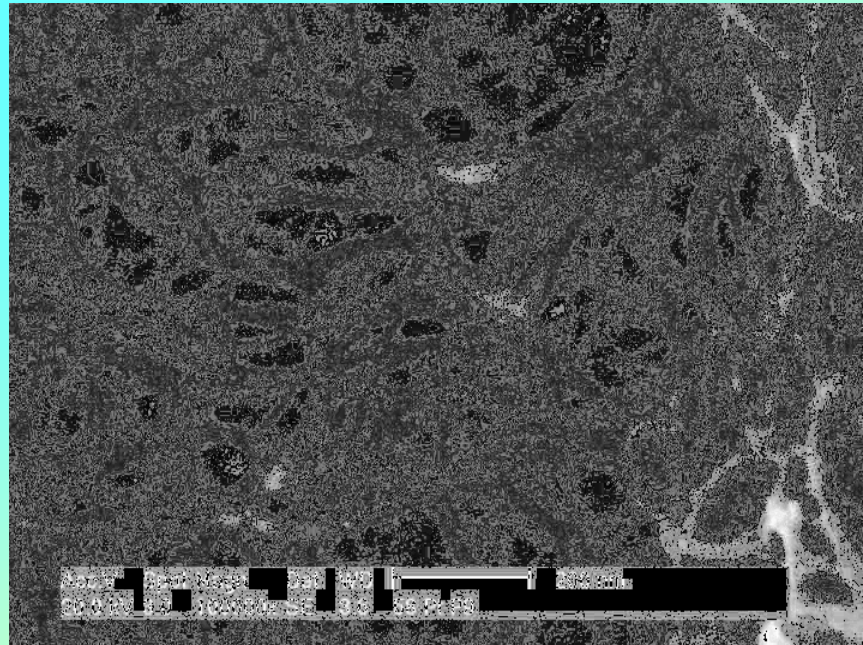
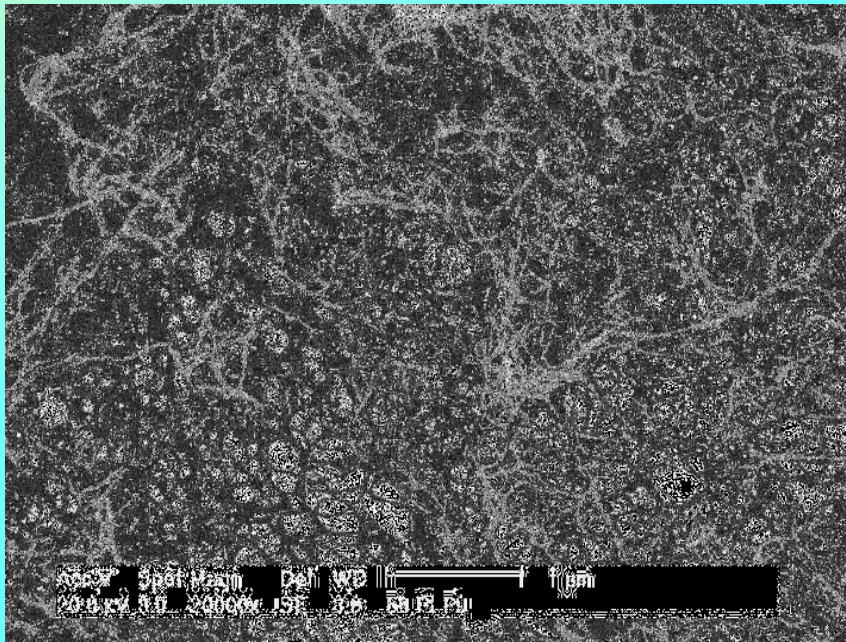
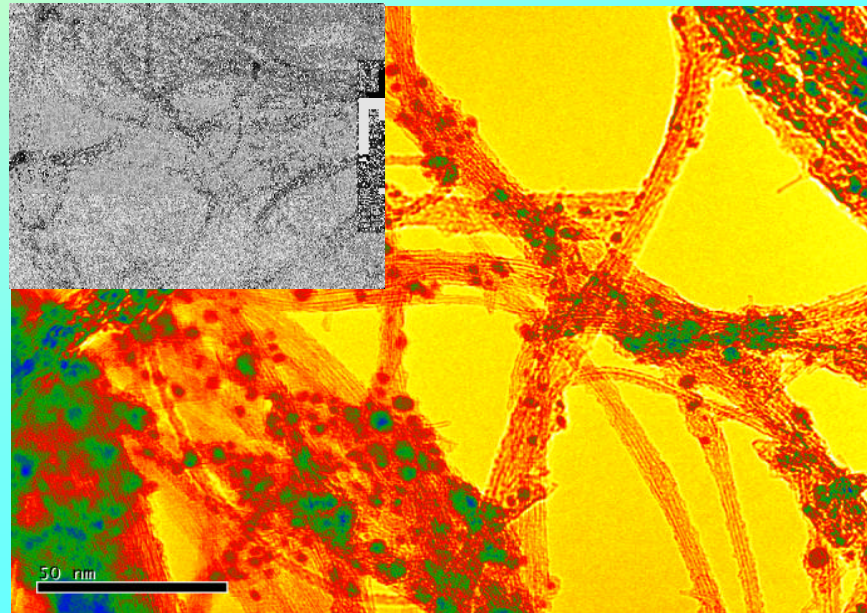
## Levels of GSH in BAL of C57BL/6J mice 7 days after exposure to partially purified SWCNT.



Partially purified SWCNT (2.0 mg/kg b.w.). \*p<0.05 vs. control

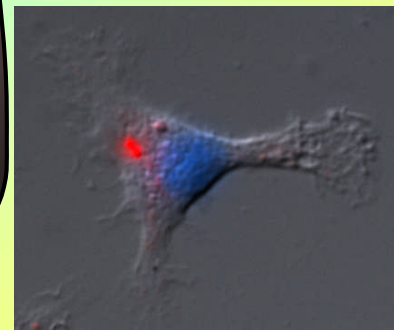
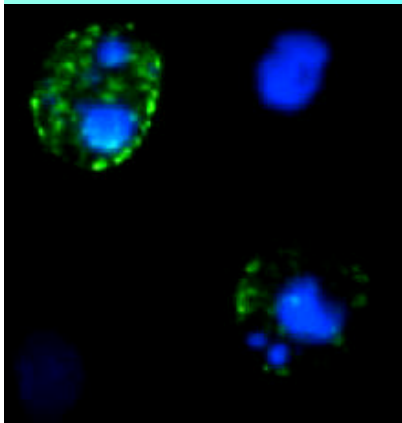
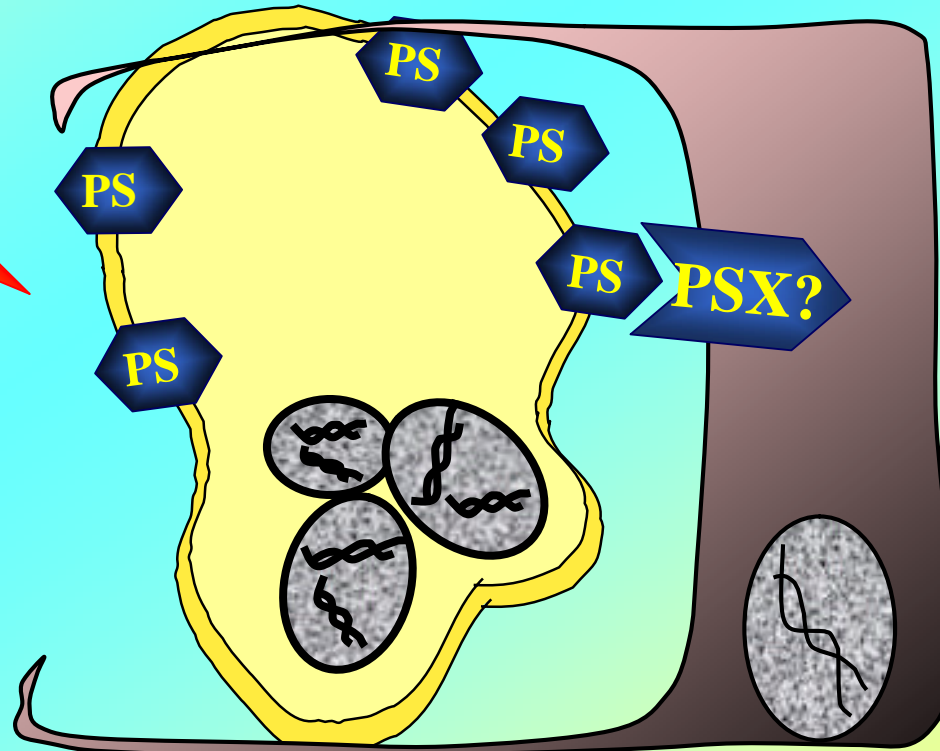
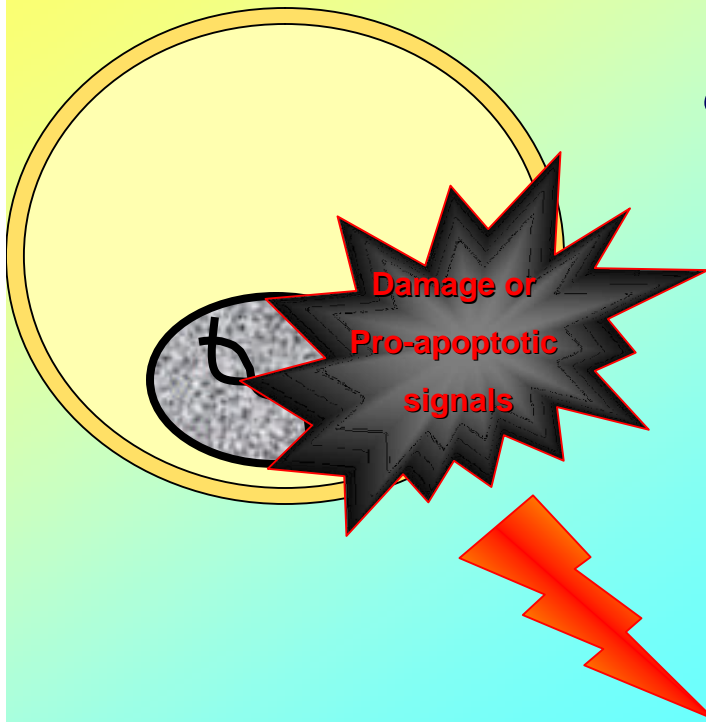


**Single walled carbon  
nanotubes  
(SWCNT) aggregate and  
form  
ropes, bundles, and  
bird's nests**

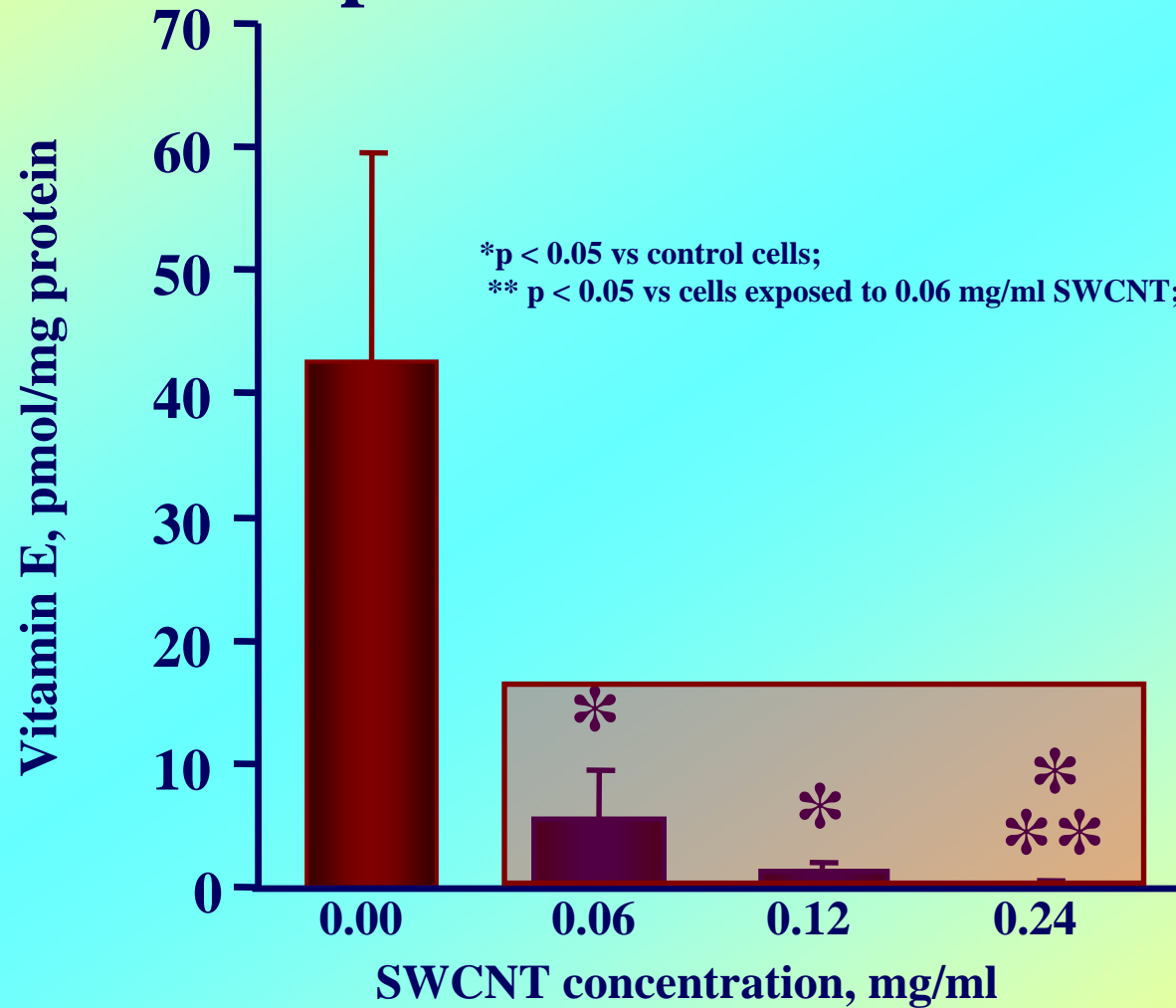




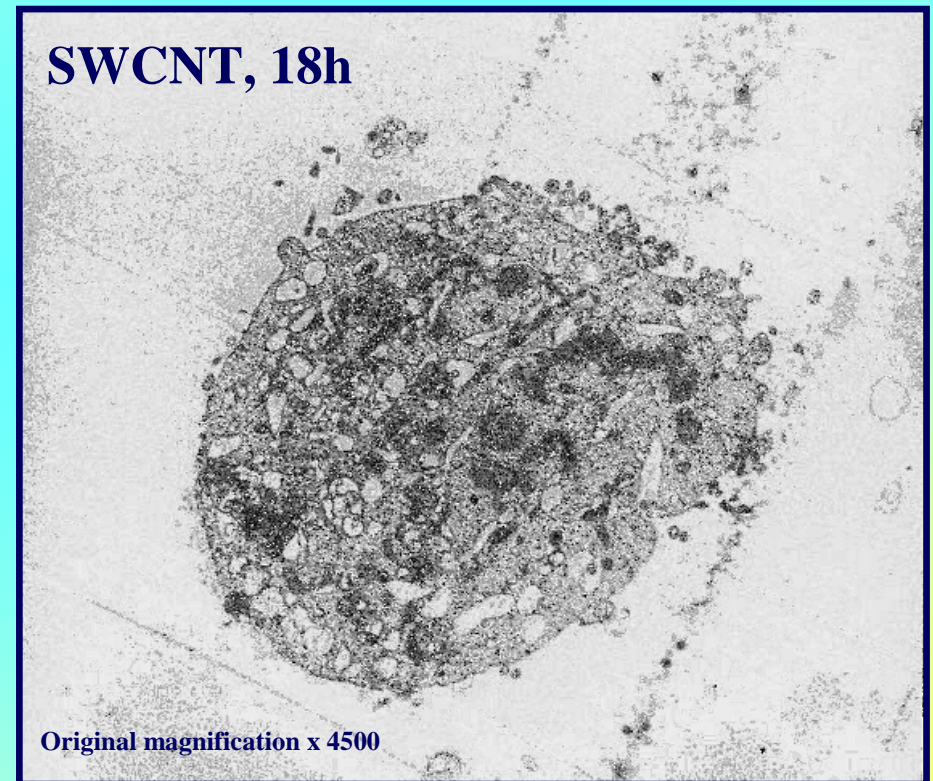
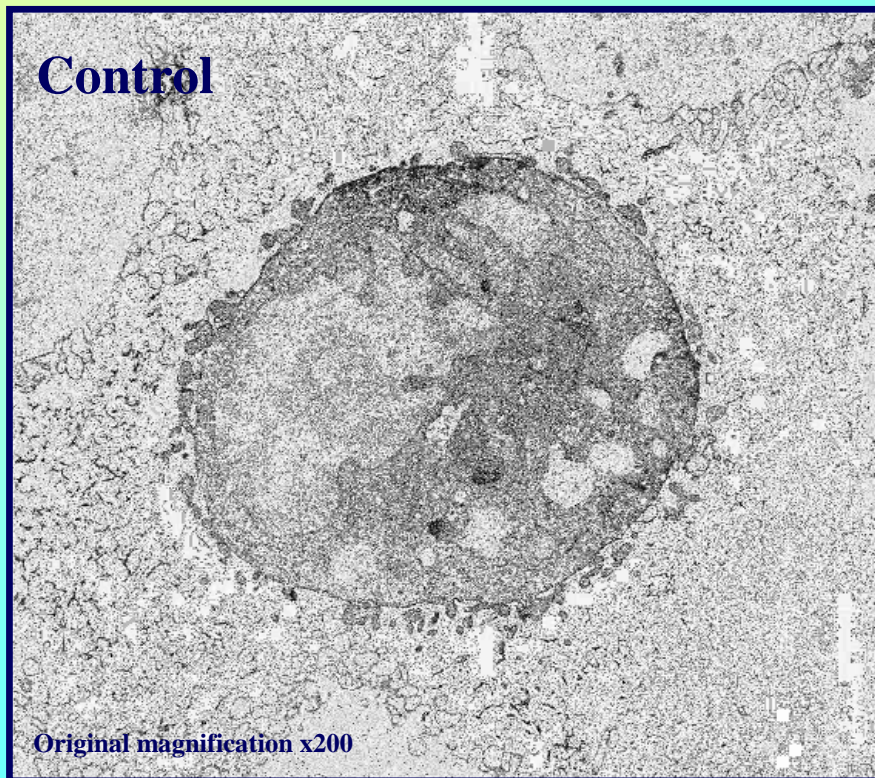
# Phosphatidylserine (PS) as an “eat-me” signal in phagocytosis of apoptotic cells



# Vitamin E Level in BEAS-2B Cells Following Exposure to SWCNT



# **Electron Microscopy of BEAS-2B Cells Exposed to SWCNT**



**SWCNT, 0.24 mg/ml**